

RESEARCH

Open Access



Prebiotic/probiotic supplementation resulted in reduced visceral fat and mRNA expression associated with adipose tissue inflammation, systemic inflammation, and chronic disease risk

Brian K. McFarlin^{1,2*}, Elizabeth A. Tanner^{1,2}, David W. Hill¹ and Jakob L. Vingren^{1,2}

Abstract

Background: Prebiotic/probiotic supplementation represents a viable option for addressing elevated systemic inflammation and chronic disease risk in overweight individuals. The purpose of this study was to determine if 90 days of prebiotic/probiotic supplementation could alter mRNA responsible for inflammation and chronic disease risk in weight-stable overweight adults. Nanostring mRNA analysis (574 plex) was used to survey targets associated with adipose tissue inflammation, systemic inflammation, and chronic disease risk. All protocols were approved by the University IRB, and participants gave written informed consent. Participants were randomly assigned to either placebo ($N = 7$; rice flour) or combined ($N = 8$) prebiotic (PreticX[®] Xylooligosaccharide; 0.8 g/day; ADIP) and probiotic (MegaDuo[®] *Bacillus subtilis* HU58 and *Bacillus coagulans* SC-208; billion CFU/day) supplementation. Participants were diverse population of healthy individuals with the exception of excess body weight. Measurements were made at baseline, 30, 60, and 90 days. Whole-body DXA scans (GE iDXA[®]; body composition) and blood 574-plex mRNA analysis (Nanostring[®]) were used to generate primary outcomes. Significance was set to $p < 0.05$ and adjusted for multiple comparisons where necessary.

Results: Compared to placebo, prebiotic/probiotic supplementation was associated with a 35% reduction in visceral adipose tissue (VAT; $p = 0.002$) but no change in body weight or overall percent body fat. Prebiotic/probiotic supplementation resulted in significant ($p < 0.05$), differential expression of 15 mRNA associated with adipose tissue inflammation (GATA3, TNFAIP6, ST2, CMKLR1, and CD9), systemic inflammation (LTF, SOCS1, and SERPING1), and/or chronic disease risk (ARG1, IL-18, CCL4, CEACAM6, ATM, CD80, and LAMP3). We also found 6 additional mRNA that had no obvious relationship to three previous biological functions (CSF1, SRC, ICAM4CD24, CD274, and CLEC6A).

Conclusion: The key findings support that 90-day prebiotic/probiotic supplementation may be associated with reduced adipose tissue inflammation, reduced systemic inflammation, and reduced chronic disease risk. Combined with the unexpected finding of reduced VAT, this intervention may have resulted in improved overall health and reduced chronic disease risk.

*Correspondence: brian.mcfarlin@unt.edu

² Department of Biological Sciences, University of North Texas, Denton, TX 76203, USA

Full list of author information is available at the end of the article



© The Author(s) 2022. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

Keywords: VAT, DXA, Body composition, Nanostring, Weight stable, Metabolically unhealthy

Background

Prebiotic/probiotic supplementation has been linked to improvement in a variety of health conditions and chronic disease states, including reduced severity of irritable bowel syndrome, reduced risk of type 2 diabetes mellitus, and reduced risk of cardiovascular disease [1]. Given that most chronic disease states have an inflammatory etiology, prebiotic/probiotic supplementation may represent a useful treatment alternative to westernized approaches. The primary mechanism of chronic disease reduction with prebiotic/probiotic is via immune system modulation and reduced peripheral/systemic inflammation [2]. While these supplements are frequently used as part of a comprehensive intervention, they are also known to exert independent effects [2, 3]. Previous research has also reported that it is possible to shift an individual from a metabolically unhealthy to a metabolically healthy phenotype without actually changing weight [4, 5]. Furthermore, the ability to uncouple body weight from metabolic health has implications not only for weight loss but also for individuals struggling to maintain weight loss [4, 5]. Typically, treatment of obesity involves a comprehensive lifestyle intervention with many components (i.e. calorie restriction, exercise intervention, behavioral treatment) [6].

Prebiotic/probiotic supplementation has known anti-inflammatory effects [7–11]; however, we speculate that reduced inflammation may have the potential to improve metabolic health, independent of weight loss. Our laboratory has previously reported that a multi-strain probiotic supplement reduced serum inflammatory cytokines and indices of dietary endotoxemia in weight stable adults [8]. Our previous study was limited to a 30-day intervention, so the present study would expand that timeline by examining changes over a 90-day period in free-living adults. The primary purpose of this study was to determine if 90 days of prebiotic/probiotic supplementation could alter mRNA responsible for inflammation and chronic disease risk in in weight stable overweight adults. A secondary purpose was to determine if there were any changes in the relative distribution of body fat.

Results

Body composition and anthropometrics

A summary of participant characteristics and baseline entry information is presented in Table 1. There was no significant percent change in total body mass, fat mass, or percent body fat in either group at baseline or

during the 90 days (Table 2). This supports the notion that the participants were weight stable. The prebiotic/probiotic group had a significant ($p = 0.002$) reduction in visceral adipose tissue (VAT) at 60 days (37%) and 90 days (35%). In comparison, placebo had a slight, nonsignificant increase in VAT during course of the study.

Overview of mRNA expression analysis

Our initial analysis identified 21 mRNA that were differentially expressed in prebiotic/probiotic compared to placebo at 30 days (Fig. 6F), 60 days (Fig. 6G), and/or 90 days (Fig. 6H). After determination of significance, we conducted a systematic review of the existing literature and Nanostring pathway annotations to group significant mRNA. Of the 21 significant mRNA, we found 15 that could be directly associated with adipose tissue inflammation (Section 3.3; Figs. 1 and 6B; GATA3, TNFAIP6, ST2, CMKLR1, and CD9), systemic inflammation (Section 3.4; Fig. 2; Fig. 6C; LTF, SOCS1, and SERPING1), and/or chronic disease risk (Section 3.5; Figs. 3 and 6D; CCL4, IL-18, ARG1, ATM, CD80, CEACAM6, and LAMP3). The remaining 6 mRNA (Section 3.6; Figs. 4 and 6E; CSF1, SRC, ICAM4, CD24, CD274, and CLEC6A) may be novel responders affected by prebiotic/probiotic supplementation but not associated with any of the above biological pathways. To allow the reader to more easily compare the changes of 15 mRNA directly linked to inflammation and chronic disease, we created Fig. 5 to demonstrate

Table 1 Participant characteristics

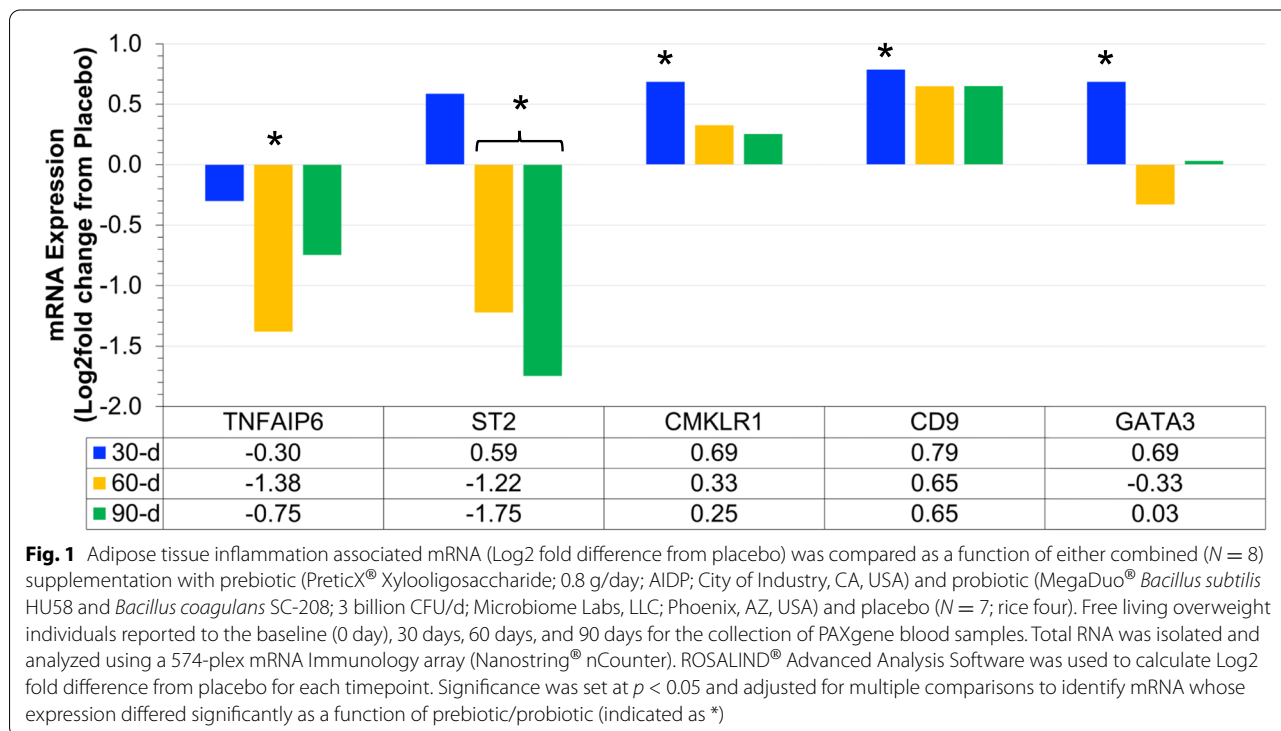
Variable	Placebo (N = 7)	Supplement (N = 8)
Sex		
(F)	5	3
(M)	2	5
Age		
(Y)	21 ± 3	22 ± 7
Height (cm)	166 ± 6	169 ± 13
BMI (kg/m ²)	27.6 ± 2.8	30.8 ± 3.5
Resting metabolic rate (kcal/day)	1603 ± 100	1871 ± 110

Probiotic Supplement (MegaDuo® Bacillus subtilis HU58 and Bacillus coagulans SC-208; 3 × 10⁹ CFU/day; Microbiome Labs, LLC; Phoenix, AZ, USA). Values represent the mean ± standard error of the mean (SEM). Variable were compared between combined supplementation with prebiotic (PreticX® Xylooligosaccharide; 0.8 g/d; ADIP; City of Industry, CA) and probiotic (MegaDuo® Bacillus subtilis HU58 and Bacillus coagulans SC-208; 3 Billion CFU/d; NuScience Trading LLC; Phoenix, AZ) and Placebo (rice flour) in free living overweight individuals over 90-d. No significant differences detected for participant characteristics between condition groups

Table 2 Participant body composition

Condition	Variable	Baseline (0 day)	30 days	60 days	90 days
Placebo (N = 7)	Mass (kg)	82.3 ± 7.8	81.8 ± 7.7	82.0 ± 7.5	82.2 ± 7.3
	Fat mass (kg)	33.1 ± 3.9	32.6 ± 3.7	33.0 ± 4.0	33.2 ± 4.0
	VAT (kg)	1.1 ± 0.4	1.1 ± 0.4	1.2 ± 0.4	1.1 ± 0.4
	VAT (Δ)		-0.02	0.07	0.02
	Body fat %	41.3 ± 1.8	41.0 ± 2.0	41.2 ± 2.1	41.4 ± 2.1
Supplement (N = 8)	Mass (kg)	93.1 ± 8.3	92.6 ± 7.3	92.8 ± 7.7	92.8 ± 7.7
	Fat mass (kg)	35.4 ± 4.0	34.6 ± 3.5	35.2 ± 3.8	35.4 ± 3.8
	VAT (kg)	1.2 ± 0.4	1.0 ± 0.2	0.8 ± 0.1	0.8 ± 0.2
	VAT (Δ)		-0.08	-0.23*†	-0.26*
	Body fat %	39.3 ± 2.6	38.9 ± 2.7	39.1 ± 2.5	39.3 ± 2.7

Values represent the mean ± standard error of the mean (SEM). Body composition variables were compared between combined supplementation with prebiotics (PreticX[®] Xylooligosaccharide; 0.8 g/day; ADIP; City of Industry, CA, USA) and probiotic (MegaDuo[®] *Bacillus subtilis* HU58 and *Bacillus coagulans* SC-208; 3 billion CFU/day; NuScience Trading LLC; Phoenix, AZ, USA) and placebo (rice flour) in free living overweight individuals at baseline (0 day), 30 days, 60 days, and 90 days of supplementation. †Represents significantly greater reduction in VAT than placebo at 60 days ($p < 0.05$). *Represents significant reduction from 30 days ($p < 0.05$)



both expression direction (i.e., positive or negative) and expression magnitude (i.e., log₂ fold difference). Additional representation of global changes is represented in Fig. 6.

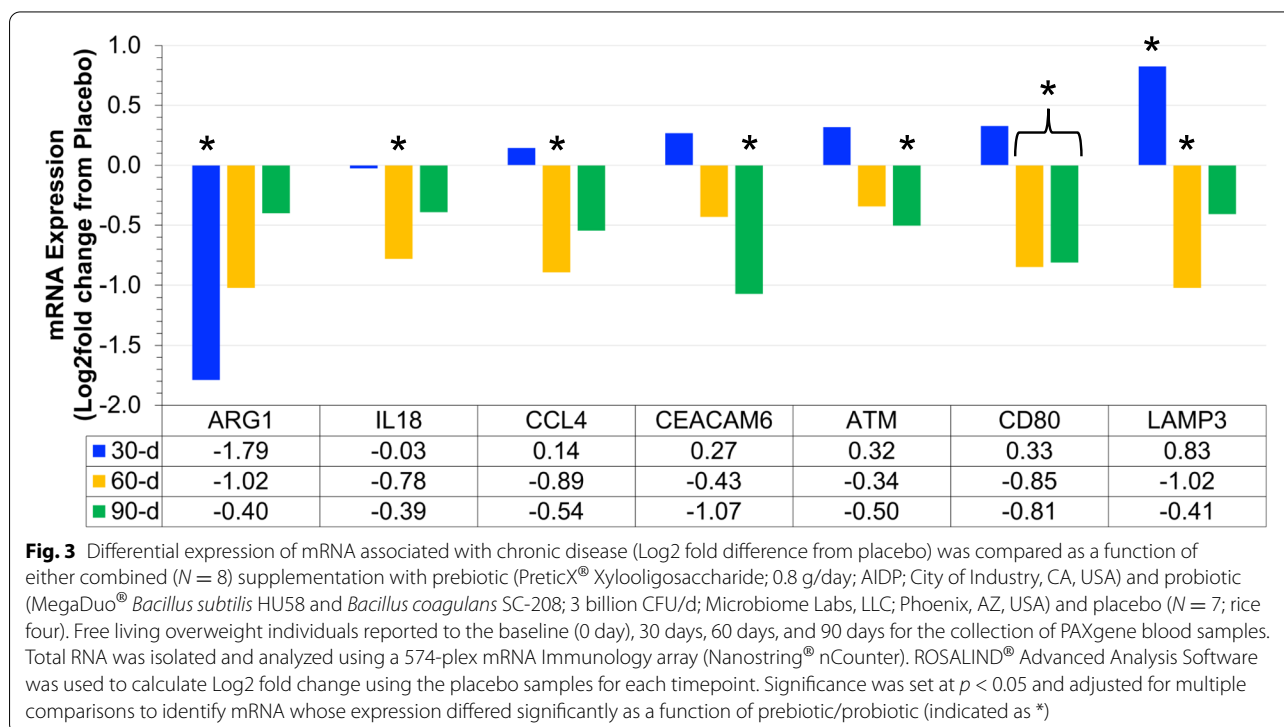
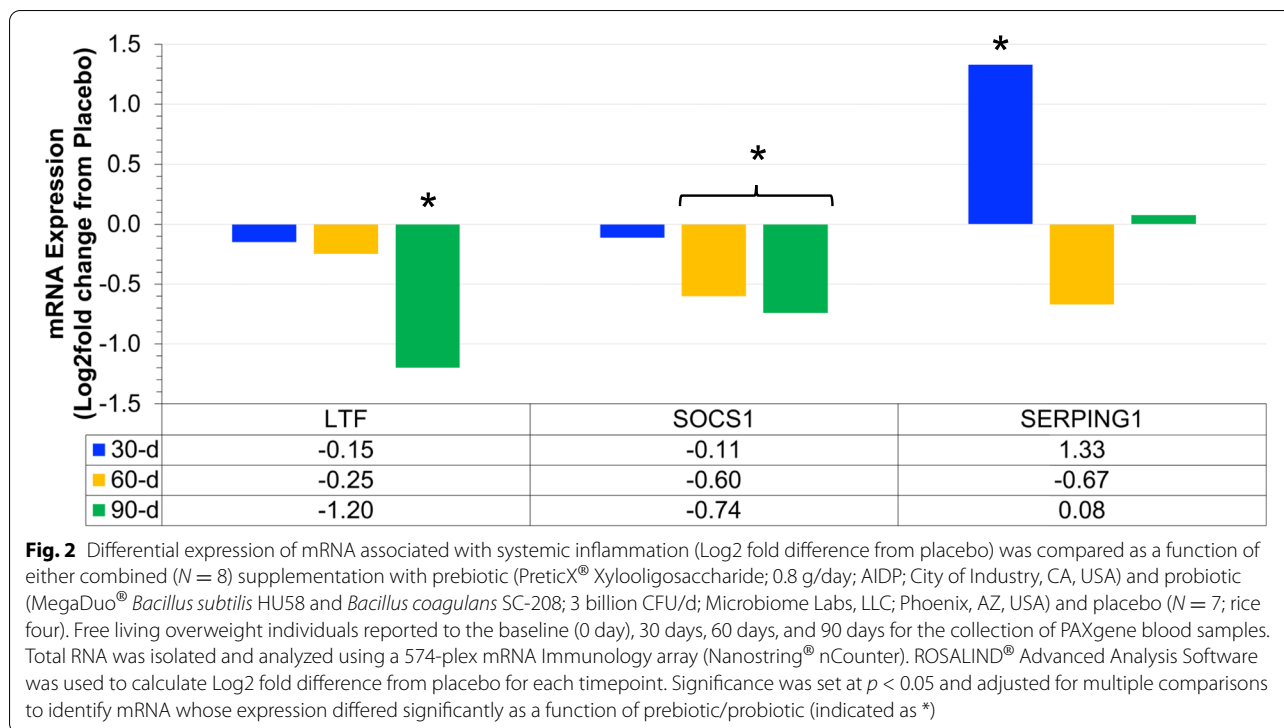
Adipose tissue inflammation mRNA expression

Prebiotic/probiotic resulted in a significant, differential mRNA expression from placebo (Figs. 1 and 6B) for GATA3 (up at 30 days; $p = 0.036$), TNFAIP6 (down at 60 days; $p = 0.003$), ST2 (down at 60 days; $p = 0.038$),

CMKLR1 (up at 30 days; $p = 0.040$), and CD9 (up at 30 days; $p = 0.028$). These 5 mRNA have been associated with adipose tissue inflammation, which is known to influence chronic, systemic inflammatory levels [12–19].

Systemic inflammation mRNA expression

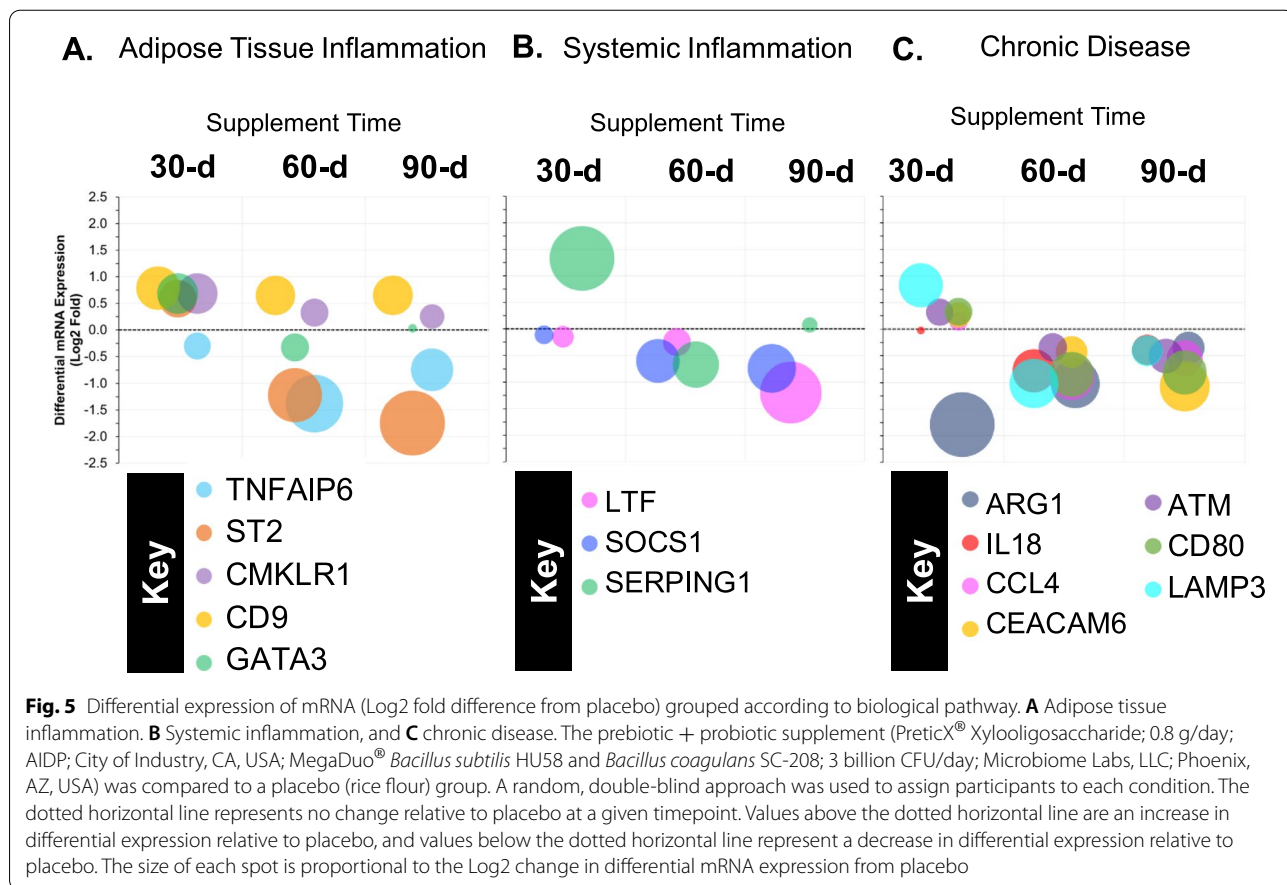
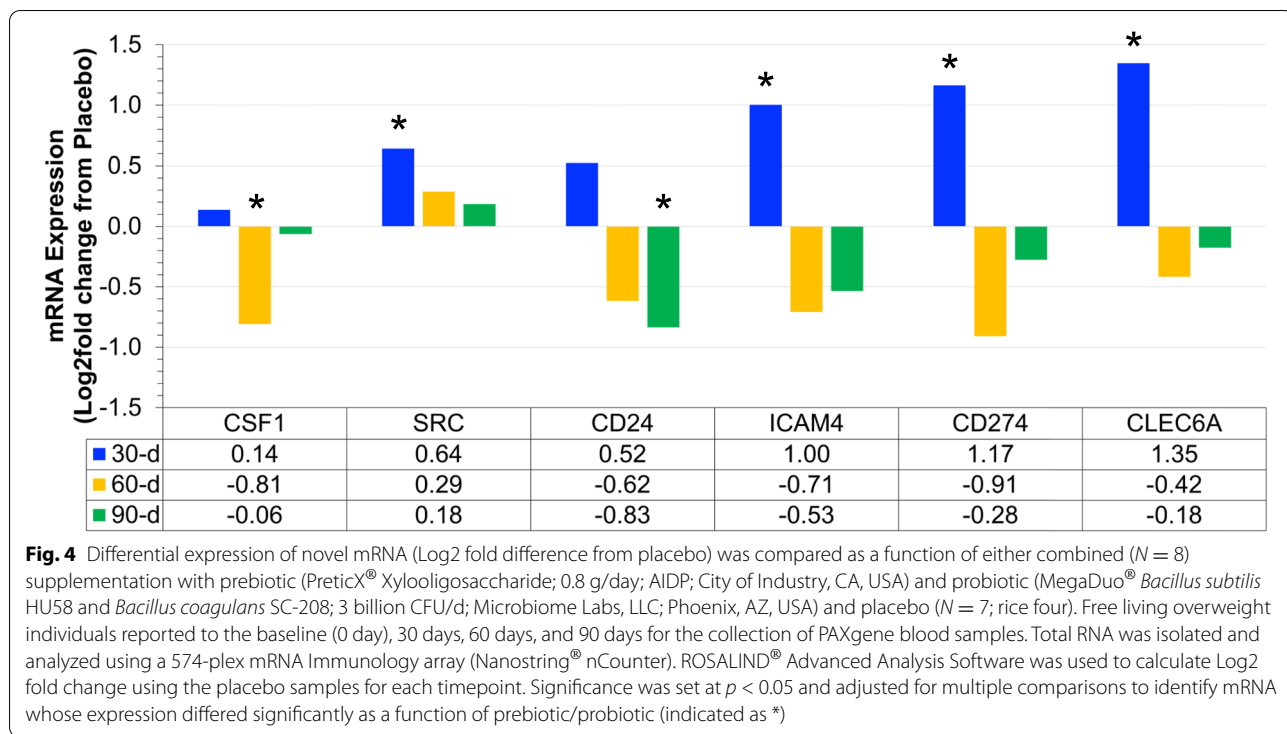
Prebiotic/probiotic resulted in a significant, differential mRNA expression from placebo (Figs. 2 and 6C) for LTF (down at 90 days; $p = 0.035$), SOCS1 (down at 60 days; $p = 0.044$; down at 90 days; $p = 0.012$), and SERPING1

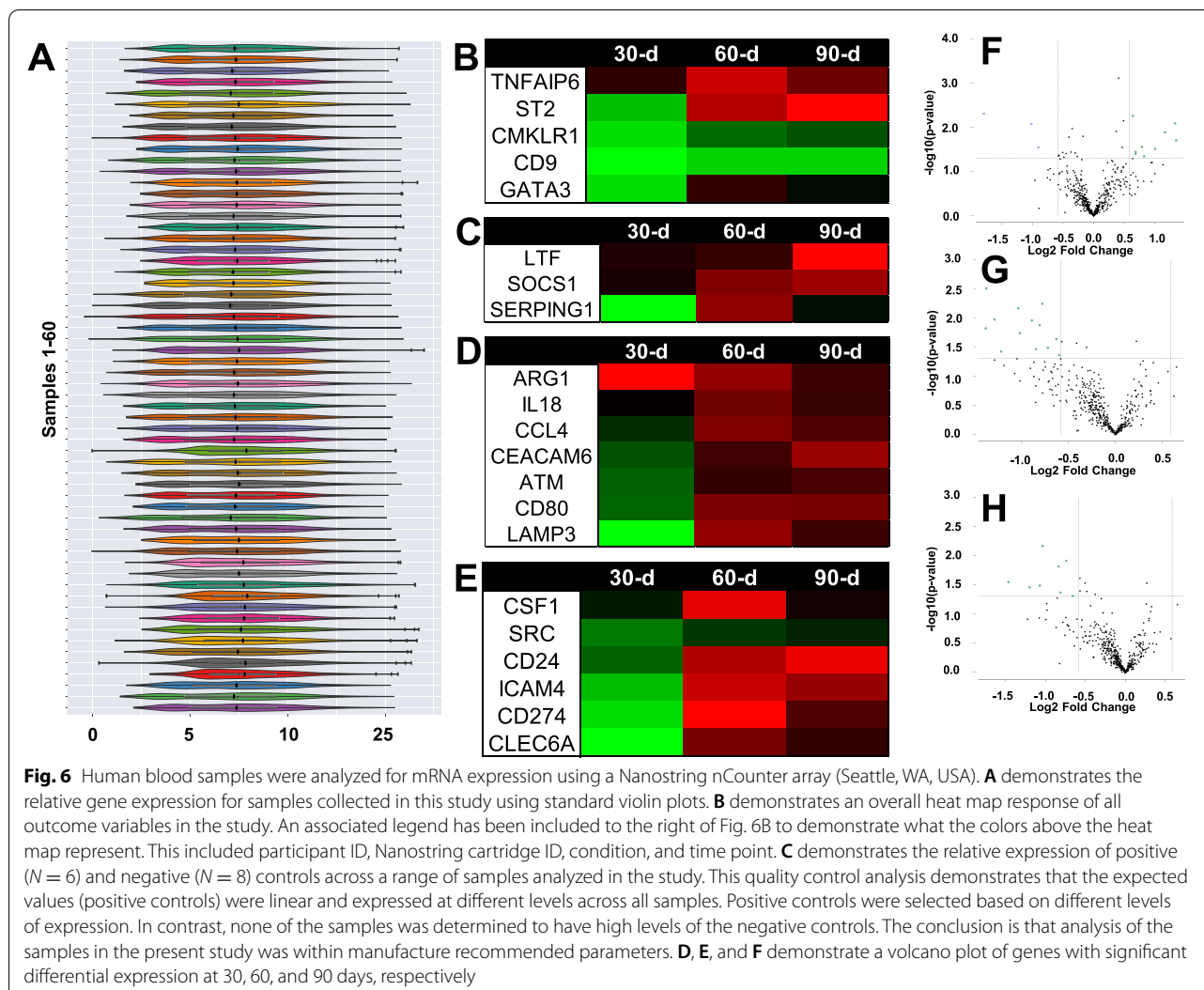


(up at 30 days; *p* = 0.008). These 3 mRNA have been previously associated with the accumulation and/or management of systemic inflammation [1, 5, 6, 20, 21].

Chronic disease mRNA expression

Prebiotic/probiotic resulted in a significant, differential mRNA expression from placebo (Figs. 3 and 6D) for





CCL4 (down at 60 days; $p = 0.011$), IL-18 (down at 60 days; $p = 0.006$), ARG1 (down at 30 days; $p = 0.005$), ATM (down at 90 days; $p = 0.030$), CD80 (down at 60 days; $p = 0.035$; down at 90 days; $p = 0.043$), CEACAM6 (down at 90 days; $p = 0.033$), and LAMP3 (up at 30 days; $p = 0.046$; down at 60 days; $p = 0.018$). These 7 mRNA have been linked to the onset and progression of cardiovascular disease, type 2 diabetes mellitus, and other chronic inflammatory diseases [22–26].

Novel/unknown mRNA expression

Prebiotic/probiotic resulted in a significant, differential mRNA expression from placebo (Figs. 4 and 6E) for CSF1 (down at 60 days; $p = 0.013$), SRC (up at 30 days; $p = 0.005$), ICAM4 (up at 30 days; $p = 0.031$), CD24 (down at 90 days; $p = 0.015$), CD274 (up at 30 days; $p = 0.013$), and CLEC6A (up at 30 days; $p = 0.020$). After reviewing

the literature and Nanostring annotated pathways, we were unable to group these 6 mRNA with others because they have no clear role in adipose tissue inflammation, systemic inflammation, or chronic disease.

Discussion

The key finding of the present study was that 90 days of prebiotic/probiotic supplementation resulted in 15 mRNA being differentially expressed from placebo. These mRNA were associated with adipose tissue inflammation, systemic inflammation, and/or chronic disease risk. Collectively, these changes may be classified as anti-inflammatory effects. We also had an unanticipated finding of an overall 36% reduction in VAT with prebiotic/probiotic supplementation compared to placebo. Previous studies have reported that similar reductions as in VAT converted obese individuals from a metabolically unhealthy

to a metabolically healthy phenotype [4, 5]. Reduced VAT combined with the significant differential mRNA changes leads us to speculate that overall health may also have been improved with prebiotic/probiotic supplementation. To our knowledge, the present study is may be first to report a potential conversion to a metabolic healthy phenotype with prebiotic/probiotic supplementation.

Adipose tissue inflammation is a well-documented side effect of weight gain and obesity [13, 15–17, 19]. Prebiotic/probiotic supplementation had pronounced impacts on differential expression of three mRNA linked to adipose tissue (GATA3, TNFAIP6, and ST2) and affected two others to a lesser degree (CMLKR1 and CD9). GATA3 has both anti-adipogenic and anti-inflammatory effects [18], while TNFAIP6 has anti-inflammatory effects [17] on peripheral adipose tissue. Interleukin 1 receptor-like 1 (ST2) promotes adipose tissue inflammation when it is ligated by IL-33 [16, 19]; ST2 expression is elevated in the adipose tissue of severely obese humans compared to normal weight individuals. Regardless, controlling propensity for weight gain and reducing adipose tissue inflammation translate to reduced chronic disease risk [12, 16]. In the present study, we found that GATA3 expression was upregulated at 30 days, while both TNFAIP6 (60 days) and ST2 (60 days and 90 days) were downregulated with prebiotic/probiotic supplementation. Considering known biological actions, prebiotic/probiotic supplementation may have contributed to a transient reduction in adipose tissue inflammation and thus improved metabolic health and reduced chronic disease risk.

We found two additional mRNA (CMKLR1 and CD9) that were also differentially expressed with prebiotic/probiotic supplementation and related to adipose tissue inflammation; however, the magnitude of change was very small compared to those described above. Chemokine-like receptor 1 (CMKLR1) is a receptor for chemerin, a potent adipokine that is elevated with obesity [13, 15]. Adipose tissue macrophages that are CD9+ contribute to adipose tissue inflammation in obese individuals [14]. Based on their biological actions, one would assume that an increase in either CMKLR1 or CD9 would not be a beneficial effect. In the present study, both factors increased to a small degree at 30 days but then rapidly declined or remained stable at 60 days and 90 days. Given the short-term response and small differential expression, it is reasonable to speculate that these effects were likely not associated with increased risk and may be a result of the body adapting to the prebiotic/probiotic supplement.

We found significant differential expression of three mRNA (LTF, SOCS1, and SERPING1) that are linked to systemic inflammation. Metabolically unhealthy obese

individuals have elevated LTF [5] and elevated SOCS1 [27]. Controlled weight loss is associated with reduced LTF [6]. Furthermore, elevated SOCS1 in obese individuals increased adipose tissue macrophage response to endotoxin, downstream pro-inflammatory cytokine production [21], and insulin resistance (1). The observed reduction in both LTF (90 days) and SOCS1 (60 days and 90 days) with prebiotic/probiotic supplementation would be considered a beneficial adaptation. SERPING1 encodes for C1 inhibitor and is negatively associated with incidence of LPS endotoxin shock in murine models [20]. Thus, SERPING1 differs from LTF and SOCS1 in that an increased expression would be considered a beneficial effect of supplement, which is what we found at 30 days in the present study. Collectively, the reduced LTF, reduced SOCS1, and increased SERPING1 may translate to a reduction in leukocyte response to LPS, reduced pro-inflammatory cytokine production, and reduced systemic inflammation.

Metabolic health is generally considered to be a precursory state that proceeds the onset and progression of chronic disease [19]. In the present study, prebiotic/probiotic supplementation was associated with change in 6 mRNA associated with reduced adipose tissue inflammation and reduced systemic inflammation. Based on the literature, we speculate that these changes support an improved metabolic health following 90 days of prebiotic/probiotic supplementation. Extending these findings further, we found significant change in 7 mRNA (CCL4, IL-18, ARG1, ATM, CD80, CEACAM6, and LAMP3) whose differential expression has been associated with various inflammation-associated chronic disease states. Elevated CCL4 [24] and IL-18 [26] result in elevated systemic inflammation and the onset and progression of type 2 diabetes mellitus. In the present study, we observed decreased differential expression of CCL4 and IL-18 at 60 days with prebiotic/probiotic supplementation, which given their associations should be associated with a reduced risk of type 2 diabetes mellitus.

Arginase 1 (ARG1), ATM, and CD80 are elevated with various forms of cardiovascular disease including the following: vascular dysfunction [28], coronary artery disease [22], and atherosclerosis [25] in obese individuals. In the present study, we found that prebiotic/probiotic supplementation reduced ARG1 (30 days), ATM (90 days), and CD80 (60 days), which may translate to a reduced cardiovascular disease risk. It was also interesting to note that the time course of change for these three indices was progressive where ARG1 was reduced first, followed by CD80 reduction, and finally with ATM reduction. It is possible that prebiotic/probiotic supplementation resulted in a coordinated response of these three mRNA. Knowing the nature of these responses, it is important for

future studies to carefully select their mRNA biomarkers based on the length of the study. Specifically, ARG1 may be better for short-term studies (< 30 days), while ATM may be better for long-term studies (> 90 days).

Type 2 diabetes mellitus and cardiovascular disease are not the only chronic disease states associated with elevated systemic. LAMP3 is elevated in obese mice and associated with the progression of non-alcoholic fatty liver disease (NAFLD) [29], while CEASAM6 is elevated in Crohn disease patients and may be an index of colon inflammation [23]. In the present study, we observed significant reductions in LAMP3 (60 days) and CECAM6 (90 days). Both LAMP3 and CECAM6 were upregulated at 30 days, followed by suppression at 60 days and 90 days. This pattern of change was similar to that observed above (CMKLR1 and CD9) and may represent the biological adjustment period to starting a prebiotic/probiotic supplement. Unfortunately, we did not have a sampling interval between 30 days and 60 days, so we are unable to narrow down the length of initial adaptation in this trial. Determining the adjustment period may be the focus of future experimental design in this area.

In addition to the above significantly changed mRNA expression, we also found that the prebiotic/probiotic supplement differentially changed the expression of 6 mRNA (CSF1, SRC, ICAM4, CD24, CD274, CLEC6A) that based on the literature cannot be directly linked to either metabolic health, inflammation, and/or chronic disease. These mRNA are summarized in Fig. 4. As with any study, the present study is not without limitations. The small sample size could be considered a limitation; however, since we found significant differential expression of 21 mRNA with prebiotic/probiotic supplementation, we have at least a partial target response. In future studies with increased sample size, we will be able to narrow our RNA biomarker panel to focus on genes associated with or related to the ones that were changed in the present study. In future studies, we will also seek to link changes in mRNA and protein biomarkers. Being able to narrow down the biomarker targets represents a substantial time and cost-savings for our laboratory and others. It is important to acknowledge that it is possible we failed to identify all mRNA that were differentially expressed with prebiotic/probiotic supplementation because we may have committed a type 2 error, which will be addressed and confirmed in future studies. While the present study included a longer supplementation period than is normal in the literature, it was by no means long duration, so more research is needed to determine the stability of the observed responses in supplementation periods exceeding 90 days. The present study was not a controlled dietary intervention, and thus changes in participant dietary habits may have confounded the present

finding. It is unknown if the observed changes are stable after prebiotic/probiotic supplementation is discontinued, so future studies may also seek to establish how discontinuation of supplementation influences metabolic health, inflammation, and chronic disease risk.

Conclusion

The present study was completed to determine if 90 days of supplementation with a combination of prebiotic/probiotic could alter mRNA responsible for inflammation and chronic disease risk in weight stable overweight adults. Our key findings were that we identified 15 mRNA whose biological action was related to adipose tissue inflammation, systemic inflammation, and/or chronic disease and were improved with 90 days of prebiotic/probiotic supplementation. We identified an additional 6 mRNA that responded to prebiotic/probiotic supplementation but have not been previously linked to inflammation/chronic disease and may be considered novel outcome measures. Meta-analysis was used to confirm that the changes over time were associated with the prebiotic/probiotic supplement. The diurnal pattern of mRNA varied considerably, highlighting the importance of longer-term interventions to maximize the effectiveness of a prebiotic/probiotic supplement on various aspects of biological function. We believe the observed changes in VAT along with the 15 mRNA support the notion that inflammation and chronic disease risk can be reduced even in the absence of weight loss in free-living overweight adults. It is our hope that the key findings of the present study will provide critical information to support future interventions aimed at reducing chronic disease risk via a prebiotic/probiotic supplement.

Methods

Study participants

All the procedures described in the present study were reviewed and approved by the University of North Texas Institutional Review Board for Human Subject's Research. Participants gave verbal and written informed consent, and procedures were conducted in accordance with the latest edition of the *Declaration of Helsinki* [30]. Exclusion criteria included body mass index less than 25 kg/m², the presence of cardiovascular or metabolic disease (i.e., diabetes mellitus), current or previous (past 12 months) tobacco or e-cig user, regular user of dietary supplements that may reduce inflammation (i.e. curcumin, pomegranate polyphenols, MSM), individual consuming an atypical diet (i.e., ketogenic, paleo, vegan), use of antibiotics, probiotics, and/or anti-inflammatory medications in the previous 6 months. Participants were recruited from the DFW metroplex, which is a diverse

community with good representation of the national population.

Participant consort tracking

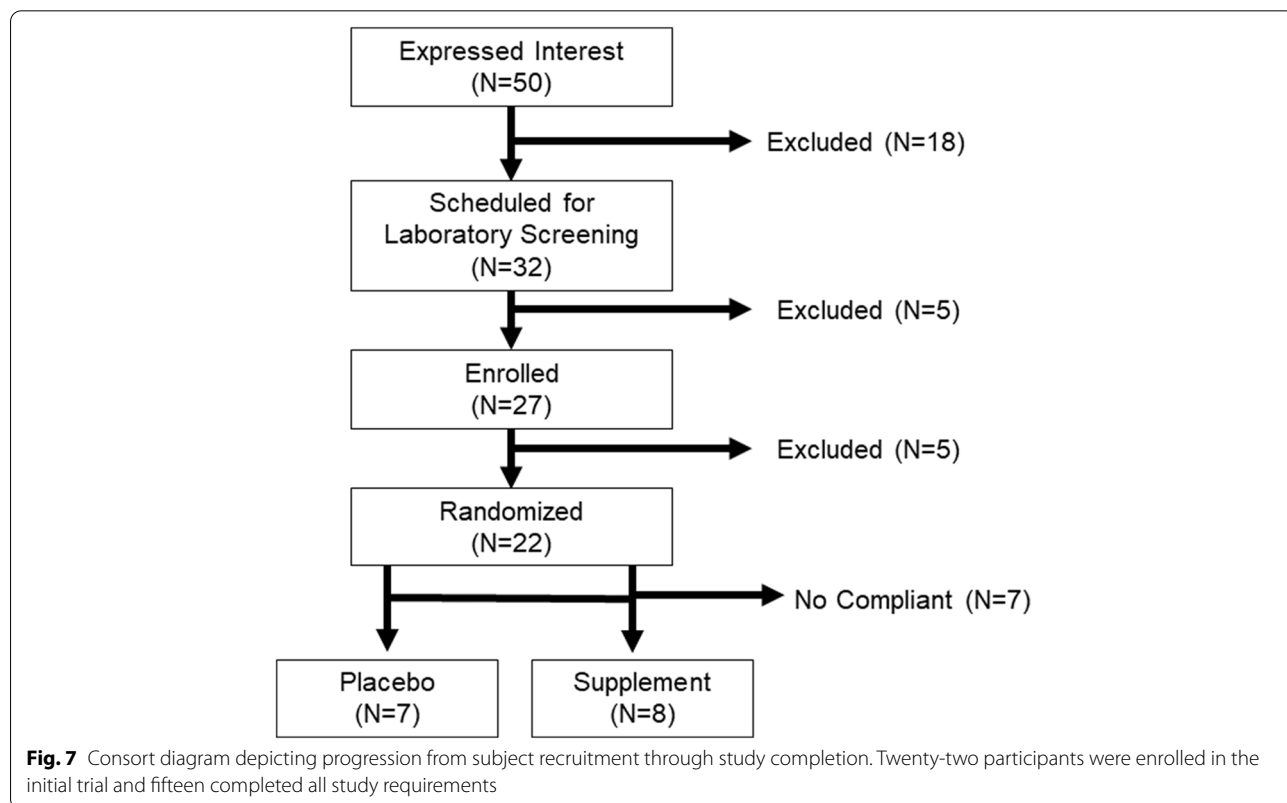
Thirty-two people provided consent for this study. Twenty-seven people met all inclusion/exclusion criteria (5 could not be scheduled further due to time constraints unrelated to the study). Twenty-two participants were randomized in an equal split ($n = 11$) to each condition, and 15 participants completed all study requirements. Seven participants were excluded for missing study appointments. Participants who were dropped from the study for missing study appointments indicated that they had a personal scheduling conflict that did not allow them to continue their participation. No participants reported side effects associated with the supplement or study participation as a reason for not continuing. A detailed breakdown of participant consort tracking can be found in Fig. 7.

Experimental design

An a priori sample size analysis was not conducted because we could not identify previously published data that would have provided data necessary for such an analysis. Despite this, we were able to estimate an approximate sample size using data from previously published studies in our laboratory [8, 31, 32]. Thus, an

objective of the present study was to establish a data set that could support future study designs in this area. We used the gold standard double-blind, placebo-controlled design to maximize applicability to future studies.

Seven of the participants who completed all study requirements were in the placebo group, and 8 were in the supplement group. The supplement consisted of a prebiotic (PreticX[®] Xylooligosaccharide, 0.8 g/day, AIDP, City of Industry, CA, USA) and a probiotic (MegaDuo[®] *Bacillus subtilis* HU58 and *Bacillus coagulans* SC-208; 3×10^9 CFU/day; Microbiome Labs, LLC; Phoenix, AZ, USA). The placebo was an equal weight of rice flour. Daily supplement (or placebo) dose was provided in three capsules, and supplementation began on day 1. Participants reported to the laboratory on four occasions: baseline (day 0), 30 days, 60 days, and 90 days. Participants were asked to bring their blister pack from the previous 30-day period to calculate compliance with the dosing requirements. When participants missed a dose during the study, they were asked to contact a member of the study staff. This information combined with the visual inspection of returned blister packs revealed that there was > 85% compliance with the supplement protocol with only two participants reporting missed doses. During the study, participants were advised not to change their diet or exercise habits to ensure a “free-living” intervention design.



At each appointment, participants provided a fasted blood sample and had their body composition and anthropometrics measured. At baseline, 30 days, and 60 days, they were provided with a 30-day supply of supplement in a blister pack. Appointments were scheduled between 0500 h and 1100 h, and participants were asked to verify that they had abstained from exercise for at least 12 h, and that they had not ingested anything other than water for at least 8 h prior to arriving at the laboratory.

Body composition and anthropometrics

Height and weight were measured using a stadiometer and digital scale respectively (SECA, USA). Participants were asked to place their back and heels against the wall, and then the top of the stadiometer was placed on their head, and height was recorded. Participants were then asked to remove shoes/socks and items from their pockets to have their weight measured. Whole-body composition was measured using a DXA scan (GE Lunar iDXA, Chicago, IL, USA). Visceral adipose tissue (VAT) was calculated (Lunar enCore CoreScan v17). The DXA machine was calibrated daily according to the manufacture recommendations. The DXA software also estimates the resting metabolic rate (RMR) using the standard Harris-Benedict equation. RMR was only used as a descriptive variable to demonstrate the two groups were not different at baseline. All body composition values were expressed as a percentage change over time compared to baseline.

Blood collection, RNA isolation, and mRNA expression analysis

Samples analysis was conducted in a manner consistent with previously published studies from our laboratory [31, 32]. Whole blood was collected from an antecubital vein in the arm into a 10 mL (2.5 mL blood + 6.5 mL PAXgene stabilizing solution) PAXgene RNA stabilizing vacutainer (PreAnalytiX; Hombrechtikon, Switzerland). Collected samples were mixed by inversion, stored at -20°C for 24 h, followed by long-term storage at -80°C until isolation and analysis. RNA was isolated as described previously [31, 32]. Briefly, total RNA was extracted using an automated (QIAcube; Qiagen, Hilden, Germany), commercially system (PAXgene Blood miRNA kit; PreAnalytiX). Isolated RNA was concentrated to 35 ng/ μL using a commercially available spin column kit (RNA Clean and Concentrator Kit, Zymo Research, Irvine, CA, USA) prior to analysis. Total RNA was analyzed using a 574-plex Human Immunology Panel v 2.0, and raw image counts (RCC files) were obtained using a Sprint Profiler (nCounter; Nanostring, Seattle, WA, USA). In addition to the targets, specific internal positive/negative controls and housekeeping mRNA (EEF1G, RPL19, PPIA, HPRT1, TBP, TUBB, POLR1B, ABCF1, SDHA, GUSB, ALAS1,

POLR2A, GAPDH, and OAZ1) were included with every RNA sample.

Statistical methods

Data were analyzed by ROSALIND[®] (<https://rosalind.onramp.bio/>) with a HyperScale architecture developed by ROSALIND, Inc. (San Diego, CA, USA). Read distribution percentages, violin plots, identity heat maps, and sample multidimensional plots were generated as part of the quality control step (Fig. 7). Normalization, fold changes, and *p*-values were calculated using criteria provided by Nanostring. ROSALIND[®] follows the nCounter[®] Advanced Analysis protocol of dividing counts within a lane by the geometric mean of the normalizer probes from the same lane. Housekeeping probes that were used for normalization were selected based on the geNorm algorithm as implemented in the NormqPCR R library [33]. Fold changes and significance score (*p*-value) were calculated using the fast method as described in the nCounter[®] Advanced Analysis 2.0 User Manual (Nanostring). Gene clustering (based on direction and type of all signals on a pathway, the position, role, and type of every gene) of differentially expressed genes was determined using the partitioning around medoids method and the fpc R library [34]. Differential expression analysis was conducted comparing supplement to placebo at three time points (30, 60, and 90 days). The clusters of mRNA generated from the three differential analyses were used to conduct a ROSALIND[®] meta-analysis to identify specific mRNA that were changed over the course of the 90-day intervention with supplement compared to placebo. Supplement mRNA were expressed as log₂ fold change from the corresponding placebo time point to normalize response to a 0 center and indicate direction of expression (i.e., up- vs. downregulated). Significant *p*-values (*p* < 0.05) were adjusted for multiple genetic comparisons using the Benjamini-Hochberg method of estimating false discovery rates [35]. Data demonstrating the relative distribution of gene expression via violin plots, heat map, and relative positive/negative control expression is presented in Fig. 6.

Abbreviations

ARG1: Arginase 1; ATM: ATM serine/threonine kinase; CD80: CD80 molecule; VAT: Visceral adipose tissue; GATA3: GATA binding protein 3; TNFAIP6: Tumor necrosis factor alpha-induced protein 6; ST2: Suppression of tumorigenicity 2; CMKLR1: Chemokine-like receptor 1; CD9: CD9 molecule; LTF: Lactotransferrin; SOCS1: Suppressor of cytokine signaling 1; SERPING1: Serpin peptidase inhibitor member 1; CCL4: Chemokine (C-C motif) ligand 4; IL-18: Interleukin-18; CEACAM6: Carcinoembryonic antigen-related cell adhesion molecule 6; LAMP3: Lysosomal-associated membrane protein 3; CSF1: Colony-stimulating factor 1; SRC: SRC proto-oncogene non-receptor tyrosine kinase; ICAM4: Inter-cellular adhesion molecule 4; CD24: CD24 molecule; CD274: CD274 molecule; CLEC6A: C-type lectin domain family 6 member A.

Acknowledgements

Not applicable

Authors' contributions

BKM designed the study, collected data, interrupted findings, and prepared manuscript. EAT collected data, interrupted findings, and prepared manuscript. DHH and JLV interrupted findings and prepared manuscript. The authors read and approved the final manuscript.

Funding

The present study was funded by a grant from physician's exclusive to UNT. The funder advised on the design of the study, but was not involved with collection, analysis, and/or interpretation of the data or the writing on the manuscript.

Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable noncommercial request.

Declarations**Ethics approval and consent to participate**

This study was approved by the Institutional Review Board of the University of North Texas, and the informed consent was obtained from each participant.

Competing interests

The present study was funded by in part by a grant to the University of North Texas from Microbiome Labs, LLC. The authors were not directly compensated and neither company was involved with the interpretation of results or the preparation of this manuscript. The authors declare that they have no competing interests.

Author details

¹Applied Physiology Laboratory, University of North Texas, Denton, TX 76203, USA. ²Department of Biological Sciences, University of North Texas, Denton, TX 76203, USA.

Received: 21 December 2021 Accepted: 14 November 2022

Published online: 28 November 2022

References

- Dumapati R, Ramatenki V, Vadija R, Vellanki S, Vuruputuri U. Structural insights into suppressor of cytokine signaling 1 protein- identification of new leads for type 2 diabetes mellitus. *J Mol Recognit*. 2018;31(7):e2706.
- Plaza-Diaz J, Ruiz-Ojeda FJ, Vilchez-Padial LM, Gil A. Evidence of the anti-inflammatory effects of probiotics and synbiotics in intestinal chronic diseases. *Nutrients*. 2017;9(6):555.
- Guandalini S, Sansotta N. Probiotics in the treatment of inflammatory bowel disease. *Adv Exp Med Biol*. 2019;1125:101–7.
- Feinberg H, Jegouzo SAF, Rex MJ, Drickamer K, Weis WI, Taylor ME. Mechanism of pathogen recognition by human dectin-2. *J Biol Chem*. 2017;292(32):13402–14.
- Jamka M, Krzyzanowska-Jankowska P, Madry E, Lisowska A, Bogdanski P, Walkowiak J. No difference in lactoferrin levels between metabolically healthy and unhealthy obese women. *Nutrients*. 2019;11(9).
- Aasbrenn M, Farup PG, Videm V. Changes in C-reactive protein, neopterin and lactoferrin differ after conservative and surgical weight loss in individuals with morbid obesity. *Sci Rep*. 2019;9(1):17695.
- Henning AL, Venable AS, Vingren JL, Hill DW, McFarlin BK. Consumption of a high-fat meal was associated with an increase in monocyte adhesion molecules, scavenger receptors, and propensity to form foam cells. *Cytometry B Clin Cytom*. 2018;94(4):606–12.
- McFarlin BK, Henning AL, Bowman EM, Gary MA, Carbajal KM. Oral spore-based probiotic supplementation was associated with reduced incidence of post-prandial dietary endotoxin, triglycerides, and disease risk biomarkers. *World J Gastrointest Pathophysiol*. 2017;8(3):117–26.
- McFarlin BK, Carpenter KC, Henning AL, Venable AS. Consumption of a high-fat breakfast on consecutive days alters preclinical biomarkers for atherosclerosis. *Eur J Clin Nutr*. 2017;71(2):239–44.
- McFarlin BK, Venable AS, Henning AL, Sampson JN, Pennel K, Vingren JL, et al. Reduced inflammatory and muscle damage biomarkers following oral supplementation with bioavailable curcumin. *BBA Clin*. 2016;5:72–8.
- Strohacker K, Breslin WL, Carpenter KC, Davidson TR, Agha NH, McFarlin BK. Moderate-intensity, premeal cycling blunts postprandial increases in monocyte cell surface CD18 and CD11a and endothelial microparticles following a high-fat meal in young adults. *Appl Physiol Nutr Metab*. 2012;37(3):530–9.
- Al-Mansoori L, Al-Jaber H, Madani AY, Mazloun NA, Agouni A, Ramanjaneya M, et al. Suppression of GATA-3 increases adipogenesis, reduces inflammation and improves insulin sensitivity in 3T3L-1 preadipocytes. *Cell Signal*. 2020;75:109735.
- Ernst MC, Haidl ID, Zuniga LA, Dranse HJ, Rourke JL, Zabel BA, et al. Disruption of the chemokine-like receptor-1 (CMKLR1) gene is associated with reduced adiposity and glucose intolerance. *Endocrinology*. 2012;153(2):672–82.
- Hill DA, Lim HW, Kim YH, Ho WY, Foong YH, Nelson VL, et al. Distinct macrophage populations direct inflammatory versus physiological changes in adipose tissue. *Proc Natl Acad Sci U S A*. 2018;115(22):E5096–E105.
- Huang C, Wang M, Ren L, Xiang L, Chen J, Li M, et al. CMKLR1 deficiency influences glucose tolerance and thermogenesis in mice on high fat diet. *Biochem Biophys Res Commun*. 2016;473(2):435–41.
- Miller AM, Purves D, McConnachie A, Asquith DL, Batty GD, Burns H, et al. Soluble ST2 associates with diabetes but not established cardiovascular risk factors: a new inflammatory pathway of relevance to diabetes? *PLoS One*. 2012;7(10):e47830.
- Niersmann C, Hauck SM, Kannenberg JM, Rohrig K, von Toerne C, Roden M, et al. Omentin-regulated proteins combine a pro-inflammatory phenotype with an anti-inflammatory counterregulation in human adipocytes: a proteomics analysis. *Diabetes Metab Res Rev*. 2019;35(1):e3074.
- Wang L, Di LJ. Wnt/beta-catenin mediates AICAR effect to increase GATA3 expression and inhibit adipogenesis. *J Biol Chem*. 2015;290(32):19458–68.
- Zeyda M, Wernly B, Demyanets S, Kaun C, Hammerle M, Hantusch B, et al. Severe obesity increases adipose tissue expression of interleukin-33 and its receptor ST2, both predominantly detectable in endothelial cells of human adipose tissue. *Int J Obes (Lond)*. 2013;37(5):658–65.
- Mejia P, Davis AE 3rd. C1 inhibitor suppresses the endotoxic activity of a wide range of lipopolysaccharides and interacts with live gram-negative bacteria. *Shock*. 2012;38(2):220–5.
- Sachithanandan N, Graham KL, Galic S, Honeyman JE, Fynch SL, Hewitt KA, et al. Macrophage deletion of SOCS1 increases sensitivity to LPS and palmitic acid and results in systemic inflammation and hepatic insulin resistance. *Diabetes*. 2011;60(8):2023–31.
- Ahmadi A, Behmanesh M, Boroumand MA, Tavallaei M. Up-regulation of MSH2, XRCC1 and ATM genes in patients with type 2 diabetes and coronary artery disease. *Diabetes Res Clin Pract*. 2015;109(3):500–6.
- Barnich N, Darfeuille-Michaud A. Abnormal CEACAM6 expression in Crohn disease patients favors gut colonization and inflammation by adherent-invasive *E. coli*. *Virulence*. 2010;1(4):281–2.
- Chang TT, Lin LY, Chen JW. A novel resolution of diabetes: C-C chemokine motif ligand 4 is a common target in different types of diabetes by protecting pancreatic islet cell and modulating inflammation. *Front Immunol*. 2021;12:650626.
- Ewing MM, Karper JC, Abdul S, de Jong RC, Peters HA, de Vries MR, et al. T-cell co-stimulation by CD28-CD80/86 and its negative regulator CTLA-4 strongly influence accelerated atherosclerosis development. *Int J Cardiol*. 2013;168(3):1965–74.
- Sharma BR, Karki R, Kanneganti TD. Role of AIM2 inflammasome in inflammatory diseases, cancer and infection. *Eur J Immunol*. 2019;49(11):1998–2011.
- Ueki K, Kadowaki T, Kahn CR. Role of suppressors of cytokine signaling SOCS-1 and SOCS-3 in hepatic steatosis and the metabolic syndrome. *Hepatol Res*. 2005;33(2):185–92.
- Bhatta A, Yao L, Xu Z, Toque HA, Chen J, Atawia AT, et al. Obesity-induced vascular dysfunction and arterial stiffening requires endothelial cell arginase 1. *Cardiovasc Res*. 2017;113(13):1664–76.

29. Liao X, Song L, Zhang L, Wang H, Tong Q, Xu J, et al. LAMP3 regulates hepatic lipid metabolism through activating PI3K/Akt pathway. *Mol Cell Endocrinol.* 2018;470:160–7.
30. World Medical A. World Medical Association Declaration of Helsinki: ethical principles for medical research involving human subjects. *JAMA.* 2013;310(20):2191–4.
31. Tanner EA, Gary MA, Michalik S, Davis AA, McFarlin BK. Optimized curcumin, pomegranate extract, and methylsulfonylmethane reduce acute, systemic inflammatory response to a half-marathon race. *Altern Ther Health Med.* 2022;28(6):72–81.
32. Tanner EA, Gary MA, Davis AA, Michalik S, McFarlin BK. Alterations in systemic inflammatory response following a half-marathon race with a combined curcumin and pomegranate supplement: a feasibility study. *J Diet Suppl.* 2021;18(5):461–77.
33. Perkins JR, Dawes JM, McMahon SB, Bennett DL, Orengo C, Kohl M. Read-qPCR and NormqPCR: R packages for the reading, quality checking and normalisation of RT-qPCR quantification cycle (Cq) data. *BMC Genomics.* 2012;13:296.
34. Hennig C. Cran-package fpc. <https://cran.r-project.org/web/packages/fpc/index.html>. 2019.
35. Ferreira JA. The Benjamini-Hochberg method in the case of discrete test statistics. *Int J Biostat.* 2007;3(1):Article 11.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

