

# Role of genetic variants in *ADIPOQ* in human eating behavior

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**Abstract** The beneficial effects of adiponectin and its negative correlation with BMI are well described. Adiponectin serum levels are altered in eating disorders such as anorexia nervosa, bulimia nervosa or binge eating. Here, we tested the hypothesis that (1) adiponectin serum levels correlate with human eating behavior factors and (2) that genetic variants of the *ADIPOQ* locus influence both serum levels and eating behavior. We analyzed 11 SNPs within *ADIPOQ* and in the 5' UTR and measured serum adiponectin levels in 1,036 individuals from the German Sorbs population. The German version of the three-factor eating questionnaire (FEV) was completed by 548 Sorbs. For replication purposes, we included an independent

replication cohort from Germany ( $N = 350$ ). In the Sorbs, we observed positive correlations of restraint with adiponectin serum levels ( $P = 0.001$ ;  $r = 0.148$ ) which, however, did not withstand adjustment for covariates ( $P = 0.083$ ;  $r = 0.077$ ). In addition, four SNPs were nominally associated with serum adiponectin levels (all  $P < 0.05$ ). Of these, two variants (rs3774261; rs1501229, all  $P < 0.05$ ) were also related to disinhibition. Furthermore, three variants were exclusively associated with hunger (rs2036373,  $P = 0.049$ ) and disinhibition (rs822396; rs864265, all  $P < 0.05$ ). However, none of these associations withstood Bonferroni corrections for multiple testing (all  $P > 9.3 \times 10^{-4}$ ). In our replication cohort, we observed similar effect directions at rs1501229 for disinhibition and hunger. A meta-analysis resulted in nominal statistical significance  $P = 0.036$  ( $Z$  score 2.086) and  $P = 0.017$  ( $Z$  score 2.366), respectively. Given the observed relationship of the SNPs with adiponectin levels and eating behavior, our data support a potential role of adiponectin in human eating behavior. Whether the relationship with eating behavior is mediated by the effects of circulating adiponectin warrants further investigations.

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## Introduction

Human circulating adiponectin is a well-described adipocytokine exerting anti-inflammatory (Yokota et al. 2000) and insulin-sensitizing effects (Yamauchi et al. 2001; Berg et al. 2001). An inverse relationship to obesity and BMI with beneficial effects for insulin sensitivity was shown (Arita et al. 1999; Cnop et al. 2003). Adiponectin serum

levels may be influenced by nutritional compounds as well as physical activity, environmental components and gender (Henneman et al. 2010; Antoniadis et al. 2009; Mantzoros et al. 2006). In particular, a 1.5-fold higher concentration in women compared to men was described (Heid 2006). Besides several other loci such as *ARL15* (ADP-ribosylation factor-like 15 gene locus) (Richards et al. 2009), the strongest genetic determinants influencing circulating adiponectin concentrations were identified within the *ADIPOQ* locus (adiponectin gene locus) (Ling et al. 2009; Heid et al. 2010; Richards et al. 2009).

Physiologic effects of adiponectin are mainly mediated by binding to one of its two receptor isoforms (AdipoR1 and AdipoR2) (Zhao et al. 2005) consequently activating signaling cascades such as adenosine monophosphate-activated protein kinase (AMPK) (Yamauchi et al. 2002), and PPAR $\alpha$  transcription factor (peroxisome proliferator-activated receptor alpha) or NF- $\kappa$ B (nuclear factor 'kappa-light-chain-enhancer' of activated B cells) (Thundiyil et al. 2012). AdipoR1 is most abundantly expressed in skeletal muscle and highly affine for globular adiponectin, while AdipoR2 binds full-length and globular adiponectin and is predominantly expressed in liver (Yamauchi et al. 2003). AdipoR1 and AdipoR2 are also expressed in human adipocytes (Rasmussen et al. 2006; Fasshauer et al. 2004; Li et al. 2007). Circulating adiponectin was shown to be involved in enhanced glucose uptake via GLUT4 (glucose transporter 4) (Ceddia et al. 2005; Mao et al. 2006), as well as in enhanced fatty acid uptake and oxidation in skeletal muscle in animal models (Tomas et al. 2002; Yoon 2006). Adiponectin-dependent AMPK activation may demonstrate a link to beneficial effects of this adipokine on metabolic and cardiovascular systems (Kahn et al. 2005). However, beside these important aspects of adiponectin in peripheral tissues, less is known about its central effects in brain. One study demonstrated that adiponectin administration stimulated the AMPK activation in the arcuate hypothalamus influencing food uptake and energy expenditure (Kubota et al. 2007). While elevated adiponectin levels were present in serum and cerebrospinal fluid during fasting state in mice, these levels were normalized after refeeding, suggesting that adiponectin might influence food intake via central mechanism in the brain (Kubota et al. 2007). One may argue whether similar central effects exist in humans (Pan et al. 2006; Kos et al. 2006). Consistent with potential central effects, human adiponectin levels were also shown to be altered in eating disorders such as anorexia nervosa, binge eating disorder and bulimia nervosa (reviewed in, e.g., Bou Khalil and El Hachem 2014). In particular, many studies demonstrated elevated adiponectin serum levels in female patients affected with anorexia nervosa (Modan-Moses et al. 2007; Pannacciulli et al. 2003; Terra et al. 2013), while binge eating disorder was related to decreased

circulating adiponectin (Monteleone et al. 2003; Carnier et al. 2012). It is worth noting that inconsistent data exist for patients suffering from bulimia nervosa (Housova et al. 2005; Tagami et al. 2004; Monteleone et al. 2003). Taken together, besides its well-known effects in terms of obesity, type 2 diabetes and related metabolic conditions adiponectin seems to be involved in food intake and energy expenditure as well as in the pathophysiology of eating disorders.

Here, we tested the hypothesis that adiponectin serum levels correlate with human eating behavior factors measured by the German version of the three-factor eating questionnaire (Fragebogen zum Essverhalten—FEV). Further, we analyzed whether genetic variation in the *ADIPOQ* locus influences adiponectin serum levels and the eating behavior factors restraint, disinhibition and hunger.

## Materials and methods

### Subjects

**The Sorbs** cohort is a self-contained population from eastern Germany which was extensively phenotyped for a wide range of anthropometric and metabolic phenotypes including weight, height, waist-to-hip-ratio (WHR) and a 75-g oral glucose tolerance test (OGTT) and standardized questionnaires for individual medical history and family histories (Veeramah et al. 2011). A total of 1,036 subjects with mean age of  $48 \pm 16$  years and mean BMI  $26.9 \pm 4.9$  kg/m<sup>2</sup> were included. A total of 548 Sorbs completed the FEV (Pudel and Westenhöfer 1989), the German version of the TFEQ (Stunkard and Messick 1985) as described elsewhere (Gast et al. 2013). Total adiponectin serum levels were measured in the Sorbs population. All subjects gave written informed consent, and the study was approved by the ethics committee of the University of Leipzig. The main characteristics of the Sorbs are summarized in Table 1.

**The replication cohort** is an independent German population described elsewhere (Gast et al. 2013). A total of 350 individuals were included in the analysis (mean age of  $27 \pm 5$  years and mean BMI of  $27.0 \pm 6.2$  kg/m<sup>2</sup>; Table 1). Phenotyping included anthropometric measurements (BMI, weight, height) and human eating behavior factors measured using the FEV (Pudel and Westenhöfer 1989). The local ethics committee of the University of Leipzig approved the study.

### Genetic analysis of the *ADIPOQ* locus

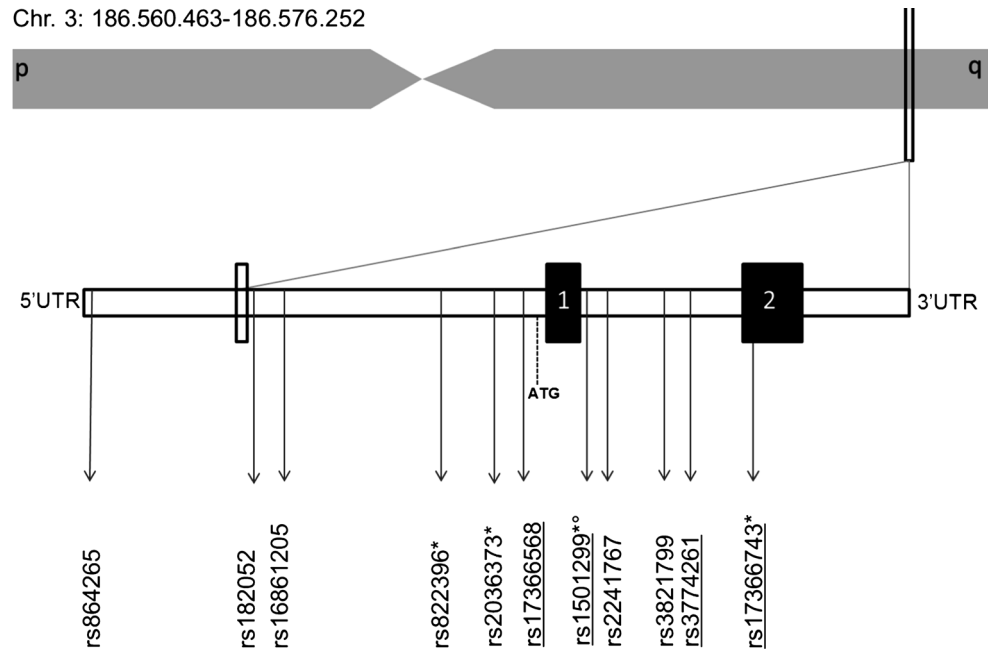
Eleven *ADIPOQ* SNPs (single nucleotide polymorphisms) within the *ADIPOQ* gene and the 5' UTR (5' untranslated

**Table 1** Main characteristics of the study populations

|                           | Sorbs      |            |            | Replication cohort |            |            |
|---------------------------|------------|------------|------------|--------------------|------------|------------|
|                           | Total      | Male       | Female     | Total              | Male       | Female     |
| N total                   | 1,036      | 418        | 618        | 350                | 189        | 152        |
| Age                       | 48 ± 16    | 48 ± 17    | 48 ± 16    | 27 ± 5             | 27 ± 5     | 27 ± 5     |
| BMI (kg/m <sup>2</sup> )  | 26.9 ± 4.9 | 27.2 ± 4.0 | 26.8 ± 5.5 | 27.0 ± 6.2         | 26.9 ± 5.8 | 27.1 ± 6.7 |
| Serum adiponectin (µg/ml) | 16.6 ± 5.5 | 14.5 ± 5.1 | 18.0 ± 5.3 | n.a.               | n.a.       | n.a.       |

Data are presented as mean ± SD

**Fig. 1** Gene structure of the *ADIPOQ* gene (not scaled) and its location on Chromosome 3. *Filled boxes* present coding exons, *unfilled* indicate non-coding exons. ATG: translation start; *asterisk* SNPs de novo genotyped in this study. Genotype data for the other SNPs were extracted from previous analysis (Tönjes et al. 2009). *Underlined* SNPs were significantly associated with adiponectin serum levels in the Sorbs. *Circle* SNPs remain significant with disinhibition and hunger after meta-analysis



region) were analyzed. Of these, genotypes for seven SNPs (rs864265; rs182052; rs16861205; rs17366568; rs2241767; rs3821799; rs3774261) were extracted from Affymetrix Genome-Wide Human SNP data (Affymetrix Inc., Santa Clara, CA, USA) earlier described by Tönjes et al. (2009). Four additional tagging variants (rs822396; rs1501229; rs2036373; rs17366743) were selected from the HapMap database ( $r^2 > 0.8$ , minor allele frequency (MAF)  $< 0.05$ ) and individually genotyped (Fig. 1). De novo genotyping of rs822396; rs1501229; rs2036373; and rs17366743 was performed using TaqMan<sup>®</sup> SNP Genotyping Assay (Applied Biosystems by Life-Technologies Carlsbad, CA, USA). Fluorescence was detected by an ABI 7500 Real-Time PCR system. All SNPs were in Hardy–Weinberg equilibrium (all  $P > 0.05$ ) except rs822396 in the Sorbs. To avoid genotyping errors, a random selection (~5 %) of

the sample was re-genotyped; all genotypes matched the initially designated genotypes. Water was used as a no template control (NTC).

#### Measurement of circulating serum adiponectin

Total adiponectin serum levels in the Sorbs were measured using a high-sensitivity human adiponectin ELISA (BioVendor; Heidelberg; Germany) using antibodies specific for human adiponectin according to manufacturers' instructions. Serum samples were collected after an overnight fast.

#### Statistics

Non-normally distributed data were logarithmically transformed to approximate a normal distribution. Linear

**Table 2** Adiponectin serum levels in T2D and obesity in the Sorbs

|                           | T2D  |            | Obesity  |            |            |
|---------------------------|--|------------|--|------------|------------|
|                           | Yes  | No         | Lean   | Overweight | Obese      |
| <i>N</i> total            | 103  | 845        | 360  | 369        | 220        |
| Serum adiponectin (µg/ml) | 15.5 ± 5.9   | 16.6 ± 5.1 | 17.5 ± 5.0                                       | 15.8 ± 5.1 | 16.1 ± 5.4 |
| <i>P</i> value            | <b>0.047<sup>a</sup></b> (adjusted for BMI: 0.330) |            | <b>4.2 × 10<sup>-8b</sup>; 0.001<sup>c</sup></b> |            |            |

Circulating adiponectin levels in µg/ml are presented as mean ± SD. *P* values were calculated using unpaired *t* tests

<sup>a</sup> T2D yes versus no; <sup>b</sup> lean versus overweight; <sup>c</sup> lean versus obese

regression models adjusted for age, gender and BMI (except for BMI) were employed to test for genetic association with BMI, eating behavior factors (restraint, disinhibition and hunger) and serum adiponectin levels. Additive model of inheritance was tested. Individuals with type 2 diabetes (T2D) were excluded from the genetic association analysis for eating behavior factors and adiponectin serum levels. Linear regression analyses adjusted for age, gender and BMI were used to assess the relationship between eating behavior and serum adiponectin levels. To correct for multiple testing, we applied Bonferroni correction as suggested at: <http://www.quantitativeskills.com/sisa/calculations/bonfer.htm> (alpha niveau 0.05; number of tests/phenotypes = 55) and lowered the significance threshold to  $P < 9.3 \times 10^{-4}$ . All *P* values  $> 9.3 \times 10^{-4}$  but  $\leq 0.05$  were considered to be of nominal statistical significance. All *P* values are provided without the Bonferroni correction. SPSS statistics version 20.0.1 (SPSS, Inc.; Chicago, IL) was used for all statistical analyses. Meta-analyses were performed using METAL (Willer et al. 2010).

## Results

### Circulating adiponectin levels in the Sorbs

As expected, adiponectin serum levels were negatively correlated with BMI ( $P = 4.5 \times 10^{-5}$ ;  $r = -0.141$ , Table 2). A strong positive correlation of serum adiponectin with the eating behavior factor restraint was observed ( $P = 0.001$ ,  $r = 0.148$ ), which, however, did not withstand adjustment for covariates such as age, gender and BMI ( $P = 0.083$ ;  $r = 0.077$ ).

### ADIPOQ SNPs associated with adiponectin serum levels and human eating behavior phenotypes

We observed nominal associations between four intra-genic SNPs (rs1501229; rs17366743; rs17366568; and

rs3774261) and circulating adiponectin levels in the Sorbs (all  $P < 0.05$ , Table 3). The strongest relationship was detected at rs3774261 (Table 3), an intronic variant with minor allele carriers conferring increased serum adiponectin levels ( $P = 0.006$ ;  $\beta = 0.693$ ).

Of these four variants, two SNPs were also nominally related to disinhibition (rs1501229 and rs3774261, Table 3). Minor allele carriers of these two variants showed both elevated serum adiponectin levels and increased disinhibition scores. In addition, we observed a third SNP (rs2036373) exerting a nominal association with hunger (Table 3). Two further variants, rs822396 and rs864265, were exclusively related to disinhibition. However, none of these associations withstood Bonferroni corrections for multiple testing (all  $P > 9.3 \times 10^{-4}$ ). No relationship between the variants and BMI was observed.

### Replication analysis in an independent German cohort

Two variants (rs3774261 and rs1501229) conferring the strongest relationships to both, adiponectin serum levels and the eating behavior factor disinhibition in the Sorbs, were taken forward to replication analyses in an independent German cohort. None of the variants were related to eating behavior factors (Table 4, all  $P > 0.05$ ). Nonetheless, we observed similar effect directions for rs1501229 as in the Sorbs showing elevated disinhibition scores in minor allele carriers. We did not find similar effect directions between the Sorbs and the replication cohort for rs3774261.

### Meta-analyses

A sample size-weighted meta-analysis for rs1501229 and rs3774261 including the results from the two study populations (Sorbs and German cohort) resulted in nominal statistical significance at rs1501229 with disinhibition and hunger (combined  $P_{\text{disinhibition}} = 0.0369$  (*Z* score 2.086);  $P_{\text{hunger}} = 0.01798$  (*Z* score 2.366) Table 5).

**Table 3** Association analysis of *ADIPOQ* genetic variants with adiponectin levels and eating behavior factors in the Sorbs

| SNP        | Genotype<br>N | Gender<br>M/F | BMI<br>kg/m <sup>2</sup> | Adiponectin<br>µg/ml     | Restraint<br>N = 548 | Disinhibition<br>N = 548 | Hunger<br>N = 547        |
|------------|---------------|---------------|--------------------------|--------------------------|----------------------|--------------------------|--------------------------|
| rs864265   | T/T N = 12    | 6/6           | 27.2 ± 4.2               | 16.44 ± 4.74             | 7.38 ± 4.08          | 2.63 ± 2.00              | 2.94 ± 1.48              |
|            | T/G N = 178   | 70/108        | 26.8 ± 4.7               | 16.75 ± 5.17             | 8.09 ± 4.48          | 4.01 ± 2.70              | 3.58 ± 2.77              |
|            | G/G N = 642   | 256/386       | 26.4 ± 4.7               | 16.56 ± 5.13             | 7.84 ± 5.03          | 4.45 ± 3.05              | 4.03 ± 2.82              |
| P value    |               |               | n.s.                     | n.s.                     | n.s.                 | <b>0.015<sup>a</sup></b> | n.s.                     |
| rs182052   | A/A N = 154   | 62/92         | 26.9 ± 4.7               | 16.18 ± 4.99             | 7.75 ± 4.75          | 3.88 ± 2.81              | 3.59 ± 2.50              |
|            | A/G N = 396   | 160/236       | 26.3 ± 4.7               | 16.88 ± 5.12             | 7.67 ± 4.83          | 4.47 ± 3.18              | 4.12 ± 3.00              |
|            | G/G N = 284   | 113/171       | 26.5 ± 4.6               | 16.43 ± 5.22             | 8.22 ± 5.04          | 4.30 ± 2.77              | 3.76 ± 2.63              |
| P value    |               |               | n.s.                     | n.s.                     | n.s.                 | n.s.                     | n.s.                     |
| rs16861205 | A/A N = 3     | 1/2           | 28.2 ± 11.8              | 17.62 ± 3.40             | 10.00 ± 0.00         | 5.00 ± 0.00              | 0.00 ± 0.00              |
|            | A/G N = 135   | 50/85         | 26.9 ± 5.3               | 16.00 ± 5.02             | 7.75 ± 4.83          | 4.55 ± 3.25              | 3.90 ± 3.22              |
|            | G/G N = 697   | 283/414       | 26.4 ± 4.5               | 16.70 ± 5.15             | 7.90 ± 4.92          | 4.27 ± 2.93              | 3.92 ± 2.71              |
| P value    |               |               | n.s.                     | n.s.                     | n.s.                 | n.s.                     | n.s.                     |
| rs17366568 | A/A N = 12    | 3/9           | 22.5 ± 3.4               | 17.52 ± 4.52             | 6.61 ± 4.26          | 3.67 ± 3.71              | 3.94 ± 2.46              |
|            | A/G N = 175   | 73/102        | 26.5 ± 4.6               | 15.81 ± 5.28             | 7.61 ± 4.83          | 4.32 ± 3.22              | 3.61 ± 2.90              |
|            | G/G N = 649   | 259/390       | 26.6 ± 4.7               | 16.78 ± 5.08             | 7.98 ± 4.93          | 4.32 ± 2.89              | 3.98 ± 2.77              |
| P value    |               |               | n.s.                     | <b>0.027<sup>a</sup></b> | n.s.                 | n.s.                     | n.s.                     |
| rs2241767  | G/G N = 2     | 0/2           | 27.2 ± 2.6               | 18.71 ± 5.08             | 12.00 ± 0.00         | 2.00 ± 0.00              | 1.00 ± 0.00              |
|            | G/A N = 86    | 37/49         | 26.0 ± 4.3               | 16.84 ± 5.89             | 8.37 ± 4.52          | 4.69 ± 3.20              | 3.76 ± 2.59              |
|            | A/A N = 747   | 298/449       | 26.6 ± 4.8               | 16.57 ± 5.03             | 7.81 ± 4.94          | 4.26 ± 2.94              | 3.91 ± 2.80              |
| P value    |               |               | n.s.                     | n.s.                     | n.s.                 | n.s.                     | n.s.                     |
| rs3821799  | T/T N = 138   | 54/84         | 26.2 ± 5.1               | 17.23 ± 5.53             | 6.99 ± 4.75          | 4.92 ± 2.95              | 4.31 ± 3.04              |
|            | T/C N = 408   | 172/236       | 26.4 ± 4.4               | 16.49 ± 4.88             | 8.17 ± 5.09          | 4.14 ± 2.88              | 3.79 ± 2.69              |
|            | C/C N = 289   | 110/179       | 26.9 ± 4.9               | 16.41 ± 5.27             | 7.87 ± 4.64          | 4.27 ± 3.09              | 3.90 ± 2.78              |
| P value    |               |               | n.s.                     | n.s.                     | n.s.                 | n.s.                     | n.s.                     |
| rs3774261  | A/A N = 94    | 36/58         | 26.0 ± 5.0               | 17.85 ± 5.20             | 7.46 ± 5.04          | 4.89 ± 2.92              | 4.17 ± 2.92              |
|            | A/G N = 396   | 164/232       | 26.4 ± 4.4               | 16.61 ± 5.00             | 8.07 ± 5.16          | 4.34 ± 2.87              | 3.92 ± 2.81              |
|            | G/G N = 347   | 136/211       | 26.8 ± 5.0               | 16.22 ± 5.21             | 7.80 ± 4.56          | 4.10 ± 3.09              | 3.81 ± 2.74              |
| P value    |               |               | n.s.                     | <b>0.006<sup>a</sup></b> | n.s.                 | <b>0.019<sup>a</sup></b> | n.s.                     |
| rs822396   | G/G N = 56    | 21/35         | 26.40 ± 4.67             | 17.57 ± 4.95             | 7.89 ± 4.4           | 3.49 ± 2.6               | 3.61 ± 2.6               |
|            | A/G N = 288   | 117/171       | 26.95 ± 4.77             | 16.71 ± 4.81             | 8.06 ± 5.0           | 4.19 ± 2.9               | 3.86 ± 2.8               |
|            | A/A N = 553   | 218/335       | 26.21 ± 4.58             | 16.48 ± 5.27             | 7.84 ± 4.9           | 4.52 ± 3.1               | 3.61 ± 2.6               |
| P value    |               |               | n.s.                     | n.s.                     | n.s.                 | <b>0.036<sup>a</sup></b> | n.s.                     |
| rs1501229  | T/T N = 70    | 30/40         | 25.60 ± 4.63             | 17.70 ± 5.12             | 7.24 ± 5.35          | 4.81 ± 2.70              | 3.78 ± 2.68              |
|            | G/T N = 387   | 154/233       | 26.55 ± 4.51             | 16.80 ± 4.90             | 8.13 ± 5.10          | 4.51 ± 2.95              | 3.96 ± 2.95              |
|            | G/G N = 434   | 172/262       | 26.52 ± 4.76             | 16.31 ± 5.29             | 7.88 ± 4.61          | 4.13 ± 3.10              | 4.52 ± 2.92              |
| P value    |               |               | n.s.                     | <b>0.010<sup>a</sup></b> | n.s.                 | <b>0.016<sup>a</sup></b> | n.s.                     |
| rs2036373  | G/G N = 2     | 1/1           | 32.30 ± 12.45            | 17.83 ± 2.53             | 8.00 ± 7.1           | 4.25 ± 3.18              | 2.00 ± 0.00              |
|            | G/T N = 75    | 38/37         | 26.87 ± 4.42             | 16.50 ± 5.99             | 7.87 ± 4.8           | 4.87 ± 3.03              | 4.67 ± 3.09              |
|            | T/T N = 811   | 313/498       | 26.43 ± 4.64             | 16.65 ± 5.01             | 7.92 ± 4.            | 4.32 ± 3.01              | 3.85 ± 2.79              |
| P value    |               |               | n.s.                     | n.s.                     | n.s.                 | n.s.                     | <b>0.049<sup>a</sup></b> |
| rs17366743 | C/C N = 0     |               |                          |                          |                      |                          |                          |
|            | C/T N = 39    | 18/21         | 27.30 ± 3.27             | 14.96 ± 4.84             | 8.91 ± 5.72          | 4.77 ± 3.52              | 3.59 ± 2.58              |
|            | T/T N = 855   | 339/516       | 26.43 ± 4.71             | 16.71 ± 5.12             | 7.88 ± 4.86          | 4.31 ± 2.98              | 3.92 ± 2.81              |
| P value    |               |               | n.s.                     | <b>0.036<sup>a</sup></b> | n.s.                 | n.s.                     | n.s.                     |

Data are presented as mean ± SD

n.s. non-significant ( $P > 0.05$ )

<sup>a</sup>  $P$  values (additive model of inheritance) were calculated using linear regression analysis, adjusted for age, gender and ln BMI

**Table 4** Replication analyses in an independent German cohort

| SNP       | Genotype<br>N | Gender<br>M/F | Age<br>years | BMI<br>kg/m <sup>2</sup> | Restraint   | Disinhibition | Hunger      |
|-----------|---------------|---------------|--------------|--------------------------|-------------|---------------|-------------|
| rs1501229 | T/T N = 36    | 23/13         | 26 ± 4.0     | 28.3 ± 6.7               | 7.51 ± 5.02 | 6.47 ± 2.86   | 6.03 ± 3.50 |
|           | G/T N = 121   | 65/56         | 27 ± 5.3     | 27.0 ± 6.6               | 5.46 ± 4.13 | 6.17 ± 3.21   | 5.82 ± 3.32 |
|           | G/G N = 156   | 87/69         | 27 ± 5.0     | 26.8 ± 5.8               | 6.37 ± 4.56 | 6.09 ± 3.21   | 5.14 ± 3.38 |
| P value   |               |               | n.s.         | n.s.                     | n.s.        | n.s.          | n.s.        |
| rs3774261 | A/A N = 56    | 33/23         | 26 ± 4.3     | 28.0 ± 6.9               | 7.53 ± 4.59 | 6.20 ± 2.73   | 5.82 ± 3.54 |
|           | A/G N = 133   | 74/59         | 27 ± 5.5     | 26.8 ± 6.2               | 5.43 ± 4.23 | 6.02 ± 3.22   | 5.62 ± 3.34 |
|           | G/G N = 123   | 67/56         | 26.5 ± 4.8   | 27.0 ± 5.9               | 6.36 ± 4.65 | 6.38 ± 3.32   | 5.34 ± 3.46 |
| P value   |               |               | n.s.         | n.s.                     | n.s.        | n.s.          | n.s.        |

Data are presented as mean ± SD. P values (additive model of inheritance) were calculated using linear regression analysis, adjusted for age, gender and ln BMI.

n.s. non-significant ( $P > 0.05$ )

**Table 5** Meta-analysis for Sorbs and German replication cohort

| SNP                  | Minor allele | Sorbs        |         |       | German cohort |         |       | Combined       |         |           |
|----------------------|--------------|--------------|---------|-------|---------------|---------|-------|----------------|---------|-----------|
|                      |              | P value      | $\beta$ | SE    | P value       | $\beta$ | SE    | P value        | Z score | Direction |
| <i>Disinhibition</i> |              |              |         |       |               |         |       |                |         |           |
| rs1501229            | T            | <b>0.016</b> | 0.429   | 0.178 | 0.781         | 0.067   | 0.239 | <b>0.03696</b> | 2.086   | ++        |
| rs3774261            | A            | <b>0.019</b> | 0.412   | 0.175 | 0.431         | -0.177  | 0.225 | 0.1762         | 1.352   | +–        |
| <i>Restraint</i>     |              |              |         |       |               |         |       |                |         |           |
| rs1501229            | T            | 0.800        | -0.078  | 0.307 | 0.852         | 0.069   | 0.369 | 0.9275         | -0.091  | –+        |
| rs3774261            | A            | 0.893        | -0.041  | 0.307 | 0.360         | 0.318   | 0.347 | 0.6483         | 0.456   | –+        |
| <i>Hunger</i>        |              |              |         |       |               |         |       |                |         |           |
| rs1501229            | T            | 0.097        | 0.296   | 0.178 | 0.084         | 0.487   | 0.281 | <b>0.01798</b> | 2.366   | ++        |
| rs3774261            | A            | 0.389        | 0.152   | 0.176 | 0.378         | 0.236   | 0.267 | 0.2217         | 1.222   | ++        |

P values were calculated based on effect sizes from linear regression model using additive inheritance model. Significant P values < 0.05 are presented in bold. All data are adjusted for age, gender and ln BMI. All analyses were standardized to the minor allele

## Discussion

The present study mainly supports the well-known relationship between genetic variants in the *ADIPOQ* gene locus and adiponectin serum levels. Beyond this, we observed two variants conferring increased adiponectin levels along with increased eating behavior scores which, however, did not withstand correction for multiple testing. Moreover, we found circulating adiponectin correlated with the eating behavior factor restraint in the German Sorbs.

### *ADIPOQ* locus and adiponectin serum levels

In the Sorbs, we found mean circulating adiponectin levels of  $16.62 \pm 5.10$   $\mu\text{g/ml}$ . In line with others describing significant differences between male and female adiponectin serum levels (Heid 2006) which is most likely caused by enriched testosterone levels in men inhibiting the secretion of high molecular weight (HMW) adiponectin from

adipocytes (Xu et al. 2005; Wang et al. 2008), we found ~19 % higher serum levels in women compared to men. Many studies described genetic variants in the *ADIPOQ* locus to be associated with reduced adiponectin levels in T2D, obesity and impaired insulin sensitivity (Vasseur 2002; Comuzzie et al. 2001; Ramya et al. 2013; Peters et al. 2013; Kadowaki 2006). Consistently, in the Sorbs, adiponectin serum levels are negatively correlated with BMI. The *ADIPOQ* gene locus was shown to be a major locus influencing plasma adiponectin levels with genome-wide significance (Heid 2006; Ling et al. 2009). Several studies identified rs17366568 upstream the transcription start site showing strongest associations to adiponectin serum concentration (Heid 2006; Peters et al. 2013; Cohen et al. 2011; Mather et al. 2012). In line with this, the same variant was nominally associated with circulating adiponectin in the Sorbs ( $P$  value = 0.027). Consistently with other studies, we observed three other markers influencing adiponectin serum levels (Ramya et al. 2013).



### *ADIPOQ* locus and eating behavior

Beside its well-described role in endocrine metabolism and cardiovascular function in peripheral tissues as well as the reported autocrine effects on adipocytes (Wu et al. 2003), adiponectin confers also central effects on energy expenditure or food intake (Kubota et al. 2007; Qi et al. 2004; Kadowaki et al. 2008). It was recently shown that adiponectin is also expressed in brain (Rodriguez-Pacheco et al. 2007; Psilopanagioti et al. 2009) and is functionally active (Rodriguez-Pacheco et al. 2007). In addition, several studies reported intracerebral injection (Hoyda et al. 2009a; Iwama et al. 2009; Park et al. 2011) and most importantly expression of adiponectin receptors in the brain (Hoyda et al. 2009b; Dadson et al. 2011). Further, it was shown that peripherally administered adiponectin is able to cross the blood–brain barrier and binds similarly to leptin neuronal targets in the hypothalamus (Thundyil et al. 2012; Qi et al. 2004; Kubota et al. 2007). In the present study, we observed several SNPs in the *ADIPOQ* gene locus which were nominally associated with human eating behavior factors such as disinhibition and hunger (Pudel and Westenhöfer 1989) as well as with adiponectin levels themselves. Minor allele carrier shows elevated adiponectin serum levels along with increased disinhibition scores which indicate the tendency to frequently overeat. It is of note, however, that none of these nominal associations withstands correction for multiple testing. Moreover, although we observed a positive correlation of restraint with adiponectin serum levels, no significant relationship of eating behavior factors and adiponectin concentrations was observed. Comparing low versus high adiponectin level groups results in significantly higher restraint scores in the high adiponectin group; however, these data do not withstand adjustment for gender (data not shown). Nevertheless, albeit non-significant, our data are in line with the hypothesis that adiponectin activates AMPK-mediated signaling via adiponectin receptor binding in the hypothalamus as demonstrated in animal models (Kubota et al. 2007; Minokoshi et al. 2008). This may result in overeating or increased hunger feelings. Since adiponectin levels can be further influenced by nutritional compounds, increased food intake may in turn serve as a positive feedback process (Mantzoros et al. 2006). However, this mechanism seems to be accompanied by decreased energy expenditure that may ultimately lead together with overeating to increased body weight (Kubota et al. 2007; Minokoshi et al. 2008). In contrast, Qi et al. (2004) reported a temporary loss in body weight after adiponectin injection into lateral-cerebral ventricles without decreased food intake in mice. The authors concluded that higher energy expenditure resulted in the weight loss, which was further supported by increased brown adipose

tissue UCP-1 (uncoupling protein-1) mRNA expression. It is still not yet clarified how adiponectin acts in the human brain, because animal studies are not fully comparable with humans and adiponectin in human cerebrospinal fluid is 1000-fold lower than serum levels (Pan et al. 2006; Kos et al. 2006). Further studies are warranted to better understand whether adiponectin acts directly through beneficial effects in peripheral tissues or indirectly by activating AMPK that may lead to altered food intake or energy expenditure.

Our study is limited at several aspects. In particular, in regard to our association results for eating behavior factors, it needs to be acknowledged that the TFEQ from Stunkard and Messick (1985) provides several restrictions. While the questionnaire identifies the three factors restraint, disinhibition and hunger, the subscale disinhibition was most consistently reported to be related with increased BMI and obesity as well as with higher energy intake (Bryant et al. 2008). It was demonstrated (Dykes et al. 2003; French et al. 1994) that disinhibition is strongly related to overeating without hunger feelings in certain situational circumstances which otherwise correlates with a high amount of food intake. Conflicting results, however, were reported for the relationship between restraint and BMI (Dykes et al. 2003; French et al. 1994), while in individuals with high restraint scores both increased and decreased energy intakes were observed (Bellisle et al. 2004; French et al. 1994). Moreover, since the questionnaire does not allow to drawing any conclusions in terms of energy intake, our data can only be interpreted in terms of eating behavior dimensions which are related to energy intake.

Therefore, our data need to be interpreted with caution. Despite the fact that the identified associations did not withstand Bonferroni correction, in concert with the well-known effects of SNPs on adiponectin concentrations, we suggest that genetic variants in *ADIPOQ* may potentially play a role in eating behavior which may be mediated via influencing the serum adiponectin levels. However, we are well aware that our data would not allow drawing these conclusions without including larger studies necessary to confirm the observed effects. Moreover, the sample size of the independent replication cohort is small, which is most likely one reason for the non-significant association results. Moreover, the small sample size may also lead to false-positive results. Since effects of gender and age on eating behavior are well recognized (Hays and Roberts 2008; Provencher et al. 2003; Jastreboff et al. 2014; Dakanalis et al. 2013), the large difference in age between the two cohorts in concert with the differential gender ratio may be one reason to explain the observed discrepancies in eating behavior scores. In addition to the reported other limitations, differences in eating behavior scores may have prevented us from identifying statistically significant SNP

effects. Further, we have no serum adiponectin levels available in the replication cohort.

Taken together, in addition to the known relationship between genetic variation in the *ADIPOQ* gene locus and adiponectin serum levels, our data suggest a potential correlation with human eating behavior factors. This may indicate potentially regulatory mechanisms in the brain in regard to beneficial effects of adiponectin. Whether the association with eating behavior is mediated by adiponectin levels or vice versa warrants further investigations.

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**Conflict of interest** The authors declare that they have no conflict of interest.

**Ethical standard** All procedures followed were in accordance with the ethical standards of the responsible committee on human experimentation (institutional and national) and with the Declaration of Helsinki 1975, as revised in 2000. Informed consent was obtained from all patients for being included in the study.

## References

- Antoniades C, Antonopoulos AS, Tousoulis D, Stefanadis C (2009) Adiponectin: from obesity to cardiovascular disease. *Obes Rev* 3:269–279. doi:10.1111/j.1467-789X.2009.00571.x
- Arita Y, Kihara S, Ouchi N, Takahashi M, Maeda K, Miyagawa J, Hotta K, Shimomura I, Nakamura T, Miyaoka K et al (1999) Paradoxical decrease of an adipose-specific protein, adiponectin, in obesity. *Biochem Biophys Res Commun* 1:79–83. doi:10.1006/bbrc.1999.0255
- Bellisle F, Clément K, Le Le Barzic M, Gall A, Guy-Grand B, Basdevant A (2004) The eating inventory and body adiposity from leanness to massive obesity: a study of 2509 adults. *Obes Res* 12:2023–2030
- Berg AH, Combs TP, Du X, Brownlee M, Scherer PE (2001) The adipocyte-secreted protein Acrp30 enhances hepatic insulin action. *Nat Med* 8:947–953. doi:10.1038/90992
- Bou Khalil R, El Hachem C (2014) Adiponectin in eating disorders. *Eat Weight Disord Stud Anorex Bulim Obes* 1:3–10. doi:10.1007/s40519-013-0094-z
- Bryant EJ, King NA, Blundell JE (2008) Disinhibition: its effects on appetite and weight regulation. *Obes Rev* 5:409–419. doi:10.1111/j.1467-789X.2007.00426.x
- Carnier J, Sanches Pde L, da Silva PL, de Piano A, Tock L, Campos RM, Corgosinho FC, Corrêa FA, Masquio D, do Nascimento CM, Oyama LM, Ernandes RH, de Mello MT, Tufik S, Dâmaso AR (2012) Obese adolescents with eating disorders: analysis of metabolic and inflammatory states. *Physiol Behav* 105(2):175–180. doi:10.1016/j.physbeh.2011.08.014
- Ceddia RB, Somwar R, Maida A, Fang X, Bikopoulos G, Sweeney G (2005) Globular adiponectin increases GLUT4 translocation and glucose uptake but reduces glycogen synthesis in rat skeletal muscle cells. *Diabetologia* 1:132–139. doi:10.1007/s00125-004-1609-y
- Cnop M, Havel PJ, Utzschneider KM, Carr DB, Sinha MK, Boyko EJ, Retzlaff BM, Knopp RH, Brunzell JD, Kahn SE (2003) Relationship of adiponectin to body fat distribution, insulin sensitivity and plasma lipoproteins: evidence for independent roles of age and sex. *Diabetologia* 46:459–469. doi:10.1007/s00125-003-1074-z
- Cohen SS, Gammon MD, North KE, Millikan RC, Lange EM, Williams SM, Zheng W, Cai Q, Long J, Smith JR et al (2011) *ADIPOQ*, *ADIPOR1*, and *ADIPOR2* polymorphisms in relation to serum adiponectin levels and BMI in black and white women. *Obesity* 10:2053–2062. doi:10.1038/oby.2010.346
- Comuzzie AG, Funahashi T, Sonnenberg G, Martin LJ, Jacob HJ, Kwitek Black AE, Maas D, Takahashi M, Kihara S, Tanaka S et al (2001) The genetic basis of plasma variation in adiponectin, a global endophenotype for obesity and the metabolic syndrome. *J Clin Endocrinol Metab* 9:4321–4325
- Dadson K, Liu Y, Sweeney G (2011) Adiponectin action: a combination of endocrine and autocrine/paracrine effects. *Front Endocrinol*. doi:10.3389/fendo.2011.00062
- Dakanalis A, Zanetti MA, Clerici M, Madeddu F, Riva G, Caccialanza R (2013) Italian version of the Dutch Eating Behavior Questionnaire. Psychometric proprieties and measurement invariance across sex, BMI-status and age. *Appetite*. doi:10.1016/j.appet.2013.08.010
- Dykes J, Brunner EJ, Martikainen PT, Wardle J (2003) Socioeconomic gradient in body size and obesity among women: the role of dietary restraint, disinhibition and hunger in the Whitehall II study. *Int J Obes*. doi:10.1038/sj.ijo.0802523
- Fasshauer M, Klein J, Kralisch S, Klier M, Lössner U, Blüher M, Paschke R (2004) Growth hormone is a positive regulator of adiponectin receptor 2 in 3T3-L1 adipocytes. *FEBS Lett* 1–3:27–32. doi:10.1016/S0014-5793(03)01525-4
- French S, Jeffery R, Wing RR (1994) Food intake and physical activity: a comparison of three measures of dieting. *Addict Behav* 4:401–409
- Gast MT, Tönjes A, Keller M, Horstmann A, Steinle N, Scholz M, Müller I, Villringer A, Stumvoll M, Kovacs P et al (2013) The role of rs2237781 within *GRM8* in eating behavior. *Brain Behav* 5:495–502. doi:10.1002/brb3.151
- Hays NP, Roberts SB (2008) Aspects of eating behaviors “disinhibition” and “restraint” are related to weight gain and BMI in women. *Obesity* 1:52–58. doi:10.1038/oby.2007.12
- Heid IM (2006) Genetic architecture of the *APM1* gene and its influence on adiponectin plasma levels and parameters of the metabolic syndrome in 1,727 healthy Caucasians. *Diabetes* 2:375–384. doi:10.2337/diabetes.55.02.06.db05-0747
- Heid IM, Henneman P, Hicks A, Coassin S, Winkler T, Aulchenko YS, Fuchsberger C, Song K, Hivert M, Waterworth DM et al (2010) Clear detection of *ADIPOQ* locus as the major gene for plasma adiponectin: results of genome-wide association analyses including 4659 European individuals. *Atherosclerosis* 2:412–420. doi:10.1016/j.atherosclerosis.2009.11.035
- Henneman P, Janssens ACJW, Zillikens MC, Frolich M, Frants RR, Oostra BA, van Duijn CM, van Dijk KW (2010) Menopause impacts the relation of plasma adiponectin levels with the



- metabolic syndrome. *J Intern Med* 4:402–409. doi:[10.1111/j.1365-2796.2009.02162.x](https://doi.org/10.1111/j.1365-2796.2009.02162.x)
- Housova J, Anderlova K, Krizová J, Haluzikova D, Kremen J, Kumstyrova T, Papezova H, Haluzik M (2005) Serum adiponectin and resistin concentrations in patients with restrictive and binge/purge form of anorexia nervosa and bulimia nervosa. *J Clin Endocrinol Metab* 3:1366–1370. doi:[10.1210/jc.2004-1364](https://doi.org/10.1210/jc.2004-1364)
- Hoyda TD, Samson WK, Ferguson AV (2009a) Adiponectin depolarizes parvocellular paraventricular nucleus neurons controlling neuroendocrine and autonomic function. *Endocrinology* 150:832–840. doi:[10.1210/en.2008-1179](https://doi.org/10.1210/en.2008-1179)
- Hoyda TD, Smith PM, Ferguson AV (2009b) Adiponectin acts in the nucleus of the solitary tract to decrease blood pressure by modulating the excitability of neuropeptide Y neurons. *Brain Res* 1256:76–84. doi:[10.1016/j.brainres.2008.12.012](https://doi.org/10.1016/j.brainres.2008.12.012)
- Iwama S, Sugimura Y, Murase T, Hiroi M, Goto M, Hayashi M, Arima H, Oiso Y (2009) Central adiponectin functions to inhibit arginine vasopressin release in conscious rats. *J Neuroendocrinol* 9:753–759. doi:[10.1111/j.1365-2826.2009.01894.x](https://doi.org/10.1111/j.1365-2826.2009.01894.x)
- Jastreboff AM, Gaiser EC, Gu P, Sinha R (2014) Sex differences in the association between dietary restraint, insulin resistance and obesity. *Eat Behav* 2:286–290. doi:[10.1016/j.eatbeh.2014.03.008](https://doi.org/10.1016/j.eatbeh.2014.03.008)
- Kadowaki T (2006) Adiponectin and adiponectin receptors in insulin resistance, diabetes, and the metabolic syndrome. *J Clin Investig* 7:1784–1792. doi:[10.1172/JCI29126](https://doi.org/10.1172/JCI29126)
- Kadowaki T, Yamauchi T, Kubota N (2008) The physiological and pathophysiological role of adiponectin and adiponectin receptors in the peripheral tissues and CNS. *FEBS Lett* 1:74–80. doi:[10.1016/j.febslet.2007.11.070](https://doi.org/10.1016/j.febslet.2007.11.070)
- Kahn BB, Alquier T, Carling D, Hardie DG (2005) AMP-activated protein kinase: ancient energy gauge provides clues to modern understanding of metabolism. *Cell Metab* 1:15–25. doi:[10.1016/j.cmet.2004.12.003](https://doi.org/10.1016/j.cmet.2004.12.003)
- Kos K, Harte AL, da Silva NF, Tonchev A, Chaldakov G, James S, Snead DR, Hoggart B, O'Hare JP, McTernan PG et al (2006) Adiponectin and resistin in human cerebrospinal fluid and expression of adiponectin receptors in the human hypothalamus. *J Clin Endocrinol Metab* 3:1129–1136. doi:[10.1210/jc.2006-1841](https://doi.org/10.1210/jc.2006-1841)
- Kubota N, Yano W, Kubota T, Yamauchi T, Itoh S, Kumagai H, Kozono H, Takamoto I, Okamoto S, Shiuchi T et al (2007) Adiponectin stimulates AMP-activated protein kinase in the hypothalamus and increases food intake. *Cell Metab* 1:55–68. doi:[10.1016/j.cmet.2007.06.003](https://doi.org/10.1016/j.cmet.2007.06.003)
- Li W, Tonelli J, Kishore P, Owen R, Goodman E, Scherer PE, Hawkins M (2007) Insulin-sensitizing effects of thiazolidinediones are not linked to adiponectin receptor expression in human fat or muscle. *AJP Endocrinol Metab* 5:E1301. doi:[10.1152/ajpendo.00312.2006](https://doi.org/10.1152/ajpendo.00312.2006)
- Ling H, Waterworth DM, Stirrabel HA, Pollin TI, Barter PJ, Kesäniemi YA, Mahley RW, McPherson R, Waeber G, Bersot TP et al (2009) Genome-wide linkage and association analyses to identify genes influencing adiponectin levels: the GEMS stud. *Obesity* 4:737–744. doi:[10.1038/oby.2008.625](https://doi.org/10.1038/oby.2008.625)
- Mantzoros CS, Williams CJ, Manson JE, Meigs JB, Hu FB (2006) Adherence to the Mediterranean dietary pattern is positively associated with plasma adiponectin concentrations in diabetic women. *Am J Clin Nutr* 84:328–335
- Mao X, Kikani CK, Riojas RA, Langlais P, Wang L, Ramos FJ, Fang Q, Christ-Roberts CY, Hong JY, Kim R et al (2006) APPL1 binds to adiponectin receptors and mediates adiponectin signalling and function. *Nat Cell Biol* 5:516–523. doi:[10.1038/ncb1404](https://doi.org/10.1038/ncb1404)
- Mather KJ, Christophi CA, Jablonski KA, Knowler WC, Goldberg RB, Kahn SE, Spector T, Dastani Z, Waterworth D, Richards JB et al (2012) Common variants in genes encoding adiponectin (ADIPOQ) and its receptors (ADIPOR1/2), adiponectin concentrations, and diabetes incidence in the Diabetes Prevention Program. *Diabet Med* 12:1579–1588. doi:[10.1111/j.1464-5491.2012.03662.x](https://doi.org/10.1111/j.1464-5491.2012.03662.x)
- Minokoshi Y, Shiuchi T, Lee S, Suzuki A, Okamoto S (2008) Role of hypothalamic AMP-kinase in food intake regulation. *Nutrition* 9:786–790. doi:[10.1016/j.nut.2008.06.002](https://doi.org/10.1016/j.nut.2008.06.002)
- Modan-Moses D, Stein D, Pariente C, Yaroslavsky A, Ram A, Faigin M, Loewenthal R, Yissachar E, Hemi R, Kanety H (2007) Modulation of adiponectin and leptin during refeeding of female anorexia nervosa patients. *J Clin Endocrinol Metab* 5:1843–1847. doi:[10.1210/jc.2006-1683](https://doi.org/10.1210/jc.2006-1683)
- Monteleone P, Fabrazzo M, Martiadis V, Fuschino A, Serritella C, Milici N, Maj M (2003) Opposite changes in circulating adiponectin in women with bulimia nervosa or binge eating disorder. *J Clin Endocrinol Metab* 11:5387–5391. doi:[10.1210/jc.2003-030956](https://doi.org/10.1210/jc.2003-030956)
- Pan W, Tu H, Kastin AJ (2006) Differential BBB interactions of three ingestive peptides: obestatin, ghrelin, and adiponectin. *Peptides* 4:911–916. doi:[10.1016/j.peptides.2005.12.014](https://doi.org/10.1016/j.peptides.2005.12.014)
- Pannacciulli N, Vettor R, Milan G, Granzotto M, Catucci A, Federspil G, de Giacomo P, Giorgino R, de Pergola G (2003) Anorexia nervosa is characterized by increased adiponectin plasma levels and reduced nonoxidative glucose metabolism. *J Clin Endocrinol Metab* 4:1748–1752. doi:[10.1210/jc.2002-021215](https://doi.org/10.1210/jc.2002-021215)
- Park M, Youn B, Zheng X, Wu D, Xu A, Sweeney G, Srivastava RK (2011) Globular adiponectin, acting via AdipoR1/APPL1, protects H9c2 Cells from hypoxia/reoxygenation-induced apoptosis. *PLoS ONE* 4:e19143. doi:[10.1371/journal.pone.0019143](https://doi.org/10.1371/journal.pone.0019143)
- Peters KE, Beilby J, Cadby G, Warrington NM, Bruce DG, Davis WA, Davis TM, Wiltshire S, Knuiman M, McQuillan BM et al (2013) A comprehensive investigation of variants in genes encoding adiponectin (ADIPOQ) and its receptors (ADIPOR1/R2), and their association with serum adiponectin, type 2 diabetes, insulin resistance and the metabolic syndrome. *BMC Med Genet* 1:15. doi:[10.1186/1471-2350-14-15](https://doi.org/10.1186/1471-2350-14-15)
- Provencher V, Drapeau V, Tremblay A, Després J, Lemieux S (2003) Eating behaviors and indexes of body composition in men and women from the Quebec family study. *Obes Res* 6:783–792
- Psilopanagioti A, Papadaki H, Kranioti EF, Alexandrides TK, Varakis JN (2009) Expression of adiponectin and adiponectin receptors in human pituitary gland and brain. *Neuroendocrinology* 1:38–47. doi:[10.1159/000151396](https://doi.org/10.1159/000151396)
- Pudel D, Westenhöfer J (1989) Fragebogen zum Eßverhalten (FEV). Handanweisung
- Qi Y, Takahashi N, Hileman SM, Patel HR, Berg AH, Pajvani UB, Scherer PE, Ahima RS (2004) Adiponectin acts in the brain to decrease body weight. *Nat Med* 5:524–529. doi:[10.1038/nm1029](https://doi.org/10.1038/nm1029)
- Ramya K, Ayyappa KA, Ghosh S, Mohan V, Radha V (2013) Genetic association of ADIPOQ gene variants with type 2 diabetes, obesity and serum adiponectin levels in south Indian population. *Gene* 2:253–262. doi:[10.1016/j.gene.2013.09.012](https://doi.org/10.1016/j.gene.2013.09.012)
- Rasmussen MS, Lihn AS, Pedersen SB, Bruun JM, Rasmussen M, Richelsen B (2006) Adiponectin receptors in human adipose tissue: effects of obesity, weight loss, and fat depots. *Obesity* 1:28–35. doi:[10.1038/oby.2006.5](https://doi.org/10.1038/oby.2006.5)
- Richards JB, Waterworth D, O'Rahilly S, Hivert M, Loos RJJ, Perry JRB, Tanaka T, Timpson NJ, Semple RK, Soranzo N et al (2009) A genome-wide association study reveals variants in ARL15 that influence adiponectin levels. *PLoS Genet* 12:e1000768. doi:[10.1371/journal.pgen.1000768](https://doi.org/10.1371/journal.pgen.1000768)
- Rodriguez-Pacheco F, Martinez-Fuentes AJ, Tovar S, Pinilla L, Tena-Sempere M, Dieguez C, Castaño JP, Malagon MM (2007) Regulation of pituitary cell function by adiponectin. *Endocrinology* 1:401–410. doi:[10.1210/en.2006-1019](https://doi.org/10.1210/en.2006-1019)

- Stunkard AJ, Messick S (1985) The three-factor eating questionnaire to measure dietary restraint, disinhibition and hunger. *J Psychosom Res* 1:71–83
- Tagami T, Satoh N, Usui T, Yamada K, Shimatsu A, Kuzuya H (2004) Adiponectin in anorexia nervosa and bulimia nervosa. *J Clin Endocrinol Metab* 4:1833–1837. doi:10.1210/jc.2003-031260
- Terra X, Auguet T, Agüera Z, Quesada IM, Orellana-Gavaldà JM, Aguilar C, Jiménez-Murcia S, Berlanga A, Guiu-Jurado E, Menchón JM et al (2013) Adipocytokine levels in women with anorexia nervosa. Relationship with weight restoration and disease duration. *Int J Eat Disord* 8:855–861. doi:10.1002/eat.22166
- Thundiyil J, Pavlovski D, Sobey CG, Arumugam TV (2012) Adiponectin receptor signalling in the brain. *Br J Pharmacol* 2:313–327. doi:10.1111/j.1476-5381.2011.01560.x
- Tomas E, Tsao TS, Saha AK, Murrey HE, Zhang CC, Itani SI, Lodish HF, Ruderman NB (2002) Enhanced muscle fat oxidation and glucose transport by ACRP30 globular domain: acetyl-CoA carboxylase inhibition and AMP-activated protein kinase activation. *Proc Natl Acad Sci* 99(25):16309–16313. doi:10.1073/pnas.222657499
- Tönjes A, Zeggini E, Kovacs P, Böttcher Y, Schleinitz D, Dietrich K, Morris AP, Enigk B, Rayner NW, Koriath M et al (2009) Association of FTO variants with BMI and fat mass in the self-contained population of Sorbs in Germany. *Eur J Hum Genet* 1:104–110. doi:10.1038/ejhg.2009.107
- Vasseur F (2002) Single-nucleotide polymorphism haplotypes in the both proximal promoter and exon 3 of the APM1 gene modulate adipocyte-secreted adiponectin hormone levels and contribute to the genetic risk for type 2 diabetes in French Caucasians. *Hum Mol Genet* 21:2607–2614. doi:10.1093/hmg/11.21.2607
- Veeramah KR, Tönjes A, Kovacs P, Gross A, Wegmann D, Geary P, Gasperikova D, Klimes I, Scholz M, Novembre J et al (2011) Genetic variation in the Sorbs of eastern Germany in the context of broader European genetic diversity. *Eur J Hum Genet* 9:995–1001. doi:10.1038/ejhg.2011.65
- Wang Y, Lam KSL, Yau M, Xu A (2008) Post-translational modifications of adiponectin: mechanisms and functional implications. *Biochem J* 3:623. doi:10.1042/BJ20071492
- Willer CJ, Li Y, Abecasis GR (2010) METAL: fast and efficient meta-analysis of genomewide association scans. *Bioinformatics* 17:2190–2191. doi:10.1093/bioinformatics/btq340
- Wu X, Motoshima H, Mahadev K, Stalker TJ, Scalia R, Goldstein BJ (2003) Involvement of AMP-activated protein kinase in glucose uptake stimulated by the globular domain of adiponectin in primary rat adipocytes. *Diabetes* 6:1355–1363. doi:10.2337/diabetes.52.6.1355
- Xu A, Chan KW, Hoo RLC, Wang Y, Tan KCB, Zhang J, Chen B, Lam MC, Tse C, Cooper GJS et al (2005) Testosterone selectively reduces the high molecular weight form of adiponectin by inhibiting its secretion from adipocytes. *J Biol Chem* 18:18073–18080. doi:10.1074/jbc.M414231200
- Yamauchi T, Kamon J, Waki H, Terauchi Y, Kubota N, Hara K, Mori Y, Ide T, Murakami K, Tsuboyama-Kasaoka N et al (2001) The fat-derived hormone adiponectin reverses insulin resistance associated with both lipotrophy and obesity. *Nat Med* 8:941–946. doi:10.1038/90984
- Yamauchi T, Kamon J, Minokoshi Y, Ito Y, Waki H, Uchida S, Yamashita S, Noda M, Kita S, Ueki K et al (2002) Adiponectin stimulates glucose utilization and fatty-acid oxidation by activating AMP-activated protein kinase. *Nat Med* 11:1288–1295. doi:10.1038/nm788
- Yamauchi T, Kamon J, Ito Y, Tsuchida A, Yokomizo T, Kita S, Sugiyama T, Miyagishi M, Hara K, Tsunoda M et al (2003) Cloning of adiponectin receptors that mediate antidiabetic metabolic effects. *Nature* 6941:762–769. doi:10.1038/nature01705
- Yokota T, Oritani K, Takahashi I, Ishikawa J, Matsuyama A, Ouchi N, Kihara S, Funahashi T, Tenner AJ, Tomiyama Y et al (2000) Adiponectin, a new member of the family of soluble defense collagens, negatively regulates the growth of myelomonocytic progenitors and the functions of macrophages. *Blood* 5:1723–1732
- Yoon MJ (2006) Adiponectin increases fatty acid oxidation in skeletal muscle cells by sequential activation of AMP-activated protein kinase, p38 mitogen-activated protein kinase, and peroxisome proliferator-activated receptor. *Diabetes* 9:2562–2570. doi:10.2337/db05-1322
- Zhao T, Hou M, Xia M, Wang Q, Zhu H, Xiao Y, Tang Z, Ma J, Ling W (2005) Globular adiponectin decreases leptin-induced tumor necrosis factor- $\alpha$  expression by murine macrophages: involvement of cAMP-PKA and MAPK pathways. *Cell Immunol* 1:19–30. doi:10.1016/j.cellimm.2005.12.002